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28 Accurate life history parameters are critical for effective fisheries management, in particular,
29 knowing the maximum life span of a species. Here we use a combination of mark-recapture and
30 genetic sampling to extend the minimum longevity of an elasmobranch species. Using this
31 approach, life span estimate of the lemon shark, *Negaprion brevirostris*, was increased
32 conservatively from 20.2 years to 37 years. This increase in longevity means higher vulnerability
33 and a longer recovery time from exploitation.

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35 Key Words: Fishery Management; Life Span; Natural Mortality; Mark-recapture; Genetic
36 Pedigree.

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38 Accurately modeling population dynamics is impossible without an understanding of how fast
39 individuals are naturally removed from the population, and how many offspring they produce
40 over a lifetime, both of which are influenced by life span (Hoenig, 1983). Life spans of long-
41 lived fishes are often underestimated, which has implications for calculating natural mortality
42 rates and intrinsic rebound potentials and can lead to mismanagement of stocks (Cailliet and
43 Andrews, 2008). If a species is longer-lived than originally modelled, for example, life-time
44 fecundity is underestimated, but instantaneous rates of natural mortality are overestimated.
45 Across shark species, population productivity is negatively correlated with maximum
46 reproductive age (Smith *et al.*, 1998), therefore an increase in the known maximum age within a
47 population suggests the stock is actually more vulnerable to overexploitation.

48 Several methods for validating age have been developed and applied to sharks and batoids
49 (elasmobranchs; Campana, 2001). Traditional age determination techniques involve analyzing
50 growth rings in vertebral centra, dorsal fin spines and caudal thorns (Brown & Gruber, 1988;
51 Cailliet *et al.*, 2006). However, considerable variability exists in calcification patterns, and thus
52 the utility of aging techniques, within and among taxonomic groups of elasmobranchs (Goldman,
53 2005). The precision (verification) and accuracy (validation) of aging techniques should be
54 assessed in all studies (see review by Cailliet *et al.*, 2006) using determinate methods such as
55 mark-recapture of known-age or chemically tagged fish or dating using lead:radium or bomb
56 radiocarbon, or through indeterminate methods such as marginal increment analysis (Beamish &
57 McFarlane, 1983; Brown & Gruber, 1988; Campana, 2001). Age estimation of older sharks
58 using band pairs is also less accurate due to a decrease in somatic growth, leading to a decrease

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59 in the rate of deposition of vertebral material (Francis *et al.*, 2007; Andrews *et al.*, 2011; Hamady
60 *et al.*, 2014). Many original life span estimates have been re-evaluated using bomb radiocarbon
61 methods and have often been revised upwards substantially (Campana, 2001; Cailliet &
62 Andrews, 2008).

63

64 The lemon shark *Negaprion brevirostris* (Poey, 1868) is a large coastal shark that is found on
65 both sides of the Atlantic Ocean and the Eastern Pacific, where it is exploited by recreational and
66 commercial fisheries (Compagno 1984; Carlson *et al.*, 2012; Shiffman & Hammerschlag, 2014).
67 Brown & Gruber (1988) used vertebral centra to age 110 *N. brevirostris* in Florida, U.S.A. and
68 Bimini, Bahamas. Annual band deposition on the vertebrae was validated through mark-
69 recapture of chemically-tagged individuals. Of the 110 animals aged, the oldest estimated was
70 20.2 years for a 226 cm pre-caudal length (L_{PC}) male. The study suffered, as many do, from low
71 sample sizes and hence the growth curve did not asymptote. Hoenig & Gruber (1990) estimated
72 that the theoretical maximum age for lemon sharks was 26 years since the largest specimens
73 from the Brown & Gruber (1988) study were considerably smaller than the maximum reported
74 size. It was hypothesized that the maximum longevity for lemon sharks had been largely
75 underestimated and could be extended using previously documented age at maturity (Brown &
76 Gruber, 1988), and an extensive genetic pedigree of offspring within the nurseries (Feldheim *et*
77 *al.*, 2014).

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78 Adult female *N. brevirostris* were aged in one of two ways: parental genotype reconstruction or
79 parental genotype reconstruction combined with tag-recapture. Parental genotype reconstruction
80 involved exhaustively sampling 0-3 year old *N. brevirostris* at Bimini (25°44'N, 79°18'W),
81 Bahamas, every year since 1993 (Feldheim *et al.*, 2014). Sampling occurred each June using
82 monofilament gillnets (180 m length, 2 m height) set perpendicular from shore. All captured *N.*
83 *brevirostris* were measured (cm) for pre-caudal length (L_{PC}), fork length (L_F) and total length
84 (L_T), sex determined, and tagged with a passive integrated transponder tag (PIT, Destron Fearing,
85 South St. Paul, MN, USA). See Gruber *et al.*, 2001, for a full description of methods. A small
86 piece of fin was removed and stored in 20% DMSO solution for genetic analysis. Individuals
87 were released alive after a brief holding period of no more than 7 days (Feldheim *et al.*, 2014).
88 DNA from all *N. brevirostris* were genotyped at 11 polymorphic microsatellite markers followed
89 by sibship and parental genotype reconstruction (for detailed methods see Feldheim *et al.*,
90 2002a; Wang, 2004; DiBattista *et al.*, 2008; Feldheim *et al.*, 2014). Genetic profiling of *N.*
91 *brevirostris* from 20 consecutive cohorts (1993-2012), showed that certain females faithfully
92 gave birth at this site for nearly two decades (Feldheim *et al.*, 2014). This enabled estimation of
93 minimum age of these individuals, with the conservative assumption being made that their first
94 genetically confirmed litter in Bimini was their first litter. Brown & Gruber (1988) estimated age
95 of maturity of 12.7 years with first parturition at 14 years, which was concordant with direct
96 observations of philopatric females of known age first returning to Bimini for parturition
97 (Feldheim *et al.*, 2014). This study, therefore, conservatively estimated the age of 19 females that
98 gave birth multiple times at Bimini by assuming that they were age 14 years the first year we
99 sampled any of their offspring, and died the last year we sampled any of their offspring.

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101 Tag recapture was also used to directly estimate age of one tagged female. Gravid females are
102 annually targeted for capture in late March and April, as they enter the Bimini lagoon as part of
103 an ongoing study of natal philopatry. Individuals were either captured by closely following the
104 shark from a skiff in shallow water (< 2 m depth), then placing a dip-net on its snout to incite it
105 to bite the mesh and become tangled, or encircling the shark with a large seine net deployed from
106 two small skiffs. Individuals were fitted with National Marine Fisheries Service (NMFS) M-type
107 dart tags (Kohler *et al.*, 1998) and tagged with electronic PIT tags unless they were already
108 tagged.

109 Parental genotype reconstruction (Feldheim *et al.*, 2014) revealed that at least 19 female *N.*
110 *brevirostris* giving birth at Bimini were older than Brown & Gruber's (1988) oldest individual
111 (Table I), including B-female 12, estimated to be at least 37 years old when last detected.

112 A mature female *N. brevirostris* with fresh mating wounds was captured using the netting
113 method on May 20th, 2014. A Vemco ® V16-6H tag was surgically implanted into the body
114 cavity (see Kessel *et al.*, 2014 for method). The shark had a broken NMFS dart tag and an intact
115 PIT tag, (ID A4D11). Records indicate that her first capture was 17 years earlier on April 18th,
116 1997, at a site less than 8 km away (Figure 1), with L_{PC} (196 cm) and L_T (252 cm), and a bulging
117 abdomen with movement of pups indicating this shark was pregnant at the time. The second
118 capture of this shark was on April 28th, 2005, and again, she was suspected to be pregnant.
119 According to the sibship pedigree (Feldheim *et al.*, 2014), a total of 65 of her pups have been

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120 sampled on a biennial basis between June 1997 and June 2013 (Table II). Brown & Gruber
121 (1988), estimated age of maturity of 12.7 years and first parturition at 14 years, which is
122 concordant with Feldheim *et al.* (2014). Based on this, A4D11 was at least 31 years old when
123 captured in 2014, and 32 years old when she was detected on multiple Vemco ® acoustic
124 receivers in Bimini waters in April, 2015 (Guttridge, T. L. pers. observation). This shark had a
125 pre-caudal length measurement of 203 cm in 2014; however, 23 cm smaller than the 20.2 year
126 old male sampled by Brown & Gruber (1988).

127 This study provides the first example of using genetic pedigree reconstruction and mark-
128 recapture to estimate the age of a chondrichthyan fish. It revises the minimum longevity of *N.*
129 *brevirostris* by nearly a factor of two from 20.2 years to 37 years, however, actual life span could
130 be longer. For example, genetic sampling of *N. brevirostris* in Bimini began in 1990; ‘B-Female
131 12’ (the oldest shark) was first detected in Bimini in 1989 with one offspring (sampled as a 1-
132 year old in 1990), so could have pupped in the years prior to sampling efforts. Further, our age
133 estimates are based on a minimum age at first parturition of 14 years. Of the six known-aged
134 sharks in Feldheim *et al.*’s (2014) genetic pedigree, the range of ages of first parturition in
135 Bimini was 14-17 years, although we cannot be sure they only gave birth in the Bimini nurseries.

136 Accurate age and life span information are an important component of any demographic
137 modeling, as maximum age and age at maturity are often used to estimate instantaneous rates of
138 natural mortality (Simpfendorfer *et al.*, 2005), as well as, intrinsic rebound potentials and
139 population doubling times (Simpfendorfer 2005, Smith *et al.*, 1998). This study was limited to
140 aging only female *N. brevirostris*. However, previous longevity studies have shown maximum

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141 ages of females are greater than those of males in numerous shark species (Carlson & Parsons,
142 1997; Driggers *et al.*, 2004). In addition, it is pertinent to note that for population modelling,
143 female life history parameters are considered more relevant as total lifetime fecundity and
144 mortality determine population numbers (Bishop *et al.*, 2006). Intrinsic rebound potential is a
145 demographic parameter that gives a population's productivity after removal of a proportion of
146 the population, and can be useful for understanding the status of exploited populations (Au *et al.*,
147 2015). Rebound potential is easily calculated for a given total mortality, needing only age at
148 maturity and natural mortality for a given species (Au *et al.*, 2015). Then *et al* (2014) concluded
149 that from over 200 direct estimates of natural mortality rates, using a maximum-age based
150 estimator performs the best among all estimators evaluated. Previous intrinsic rebound
151 calculations have incorporated the longevity estimate of 25 years for *N. brevirostris* (Smith *et al.*,
152 1998), or Hoenig's (1983) formula for natural mortality rates (which also uses the maximum age
153 of 25 years). With this updated minimum longevity, the estimated natural mortality rate
154 ($M=4.899*t_{max}^{-0.916}$) for *N. brevirostris* has decreased from $0.257yr^{-1}$ to $0.179yr^{-1}$ (Then *et al.*,
155 2014). This increase in life span could mean an increase in the time it takes an exploited
156 population to rebound back to stable numbers.

157 Although there is evidence of gene flow amongst *N. brevirostris* Northern Hemisphere
158 populations (Ashe *et al.*, 2015) and telemetry data indicating connectivity between the USA and
159 the Bahamas (unpublished data), the genes related to longevity have not yet been identified so
160 we cannot be certain that the longevity found in this study applies throughout their entire
161 distribution. This study may prove difficult to replicate in other species but highlights the

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162 advantage and importance of long-term, standardized tagging projects, and the collection and
163 analysis of genetic samples for life-history parameter research.

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