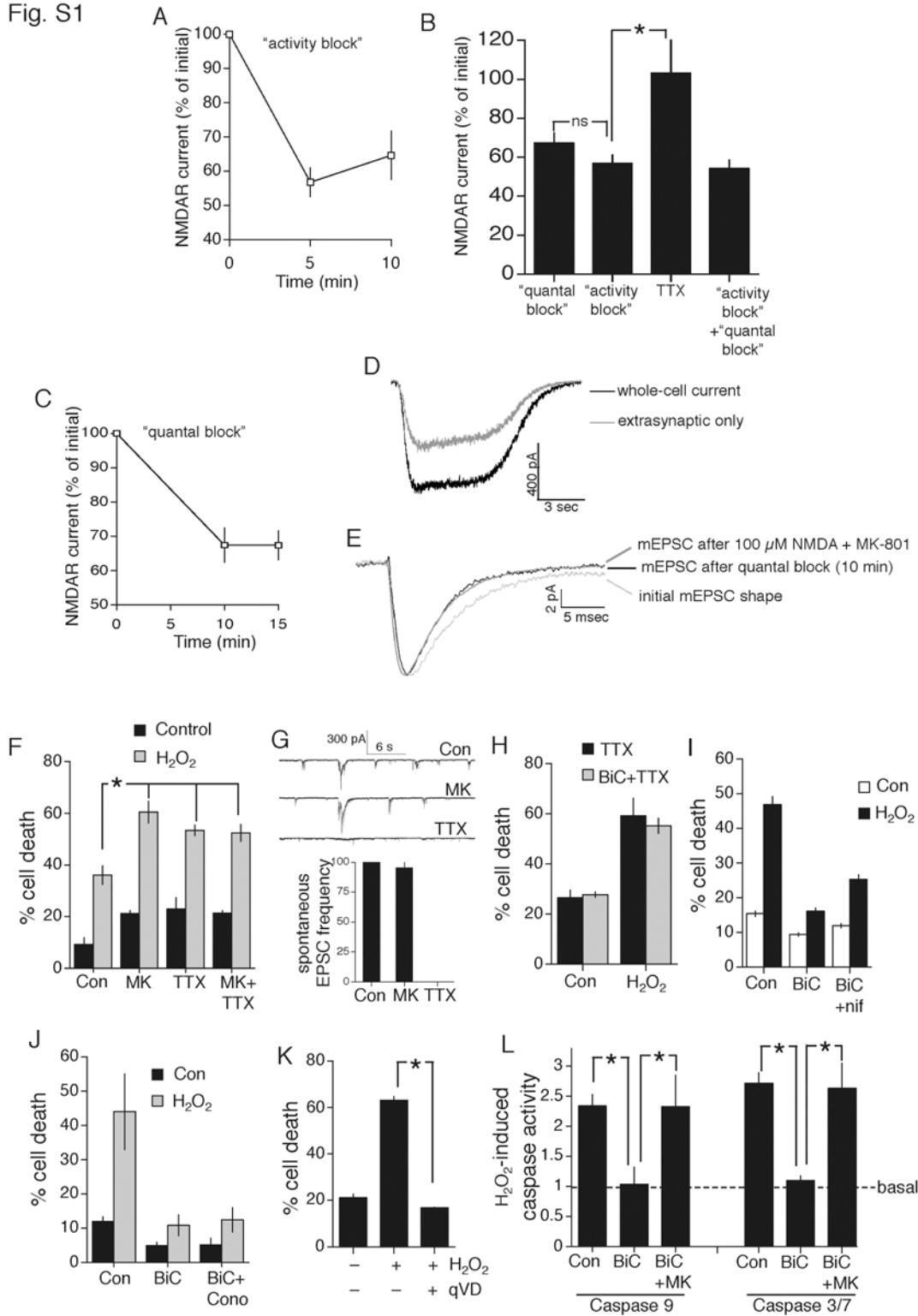


# Synaptic NMDA receptor activity boosts intrinsic antioxidant defences

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## Supplemental Figures S1-S7 and Tables T1-T2

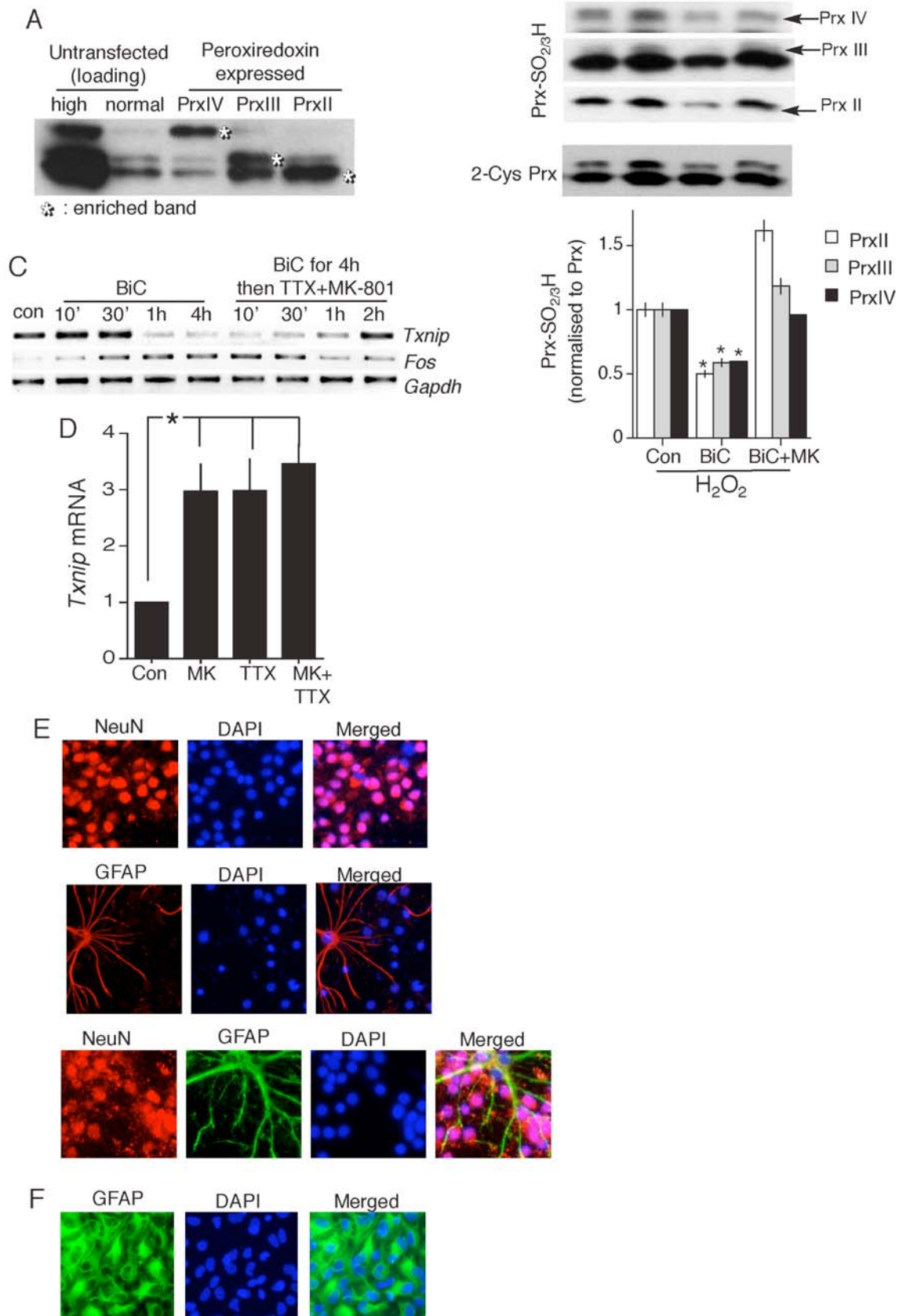
Fig. S1



**Fig. S1.** A) Loss of whole-cell current due to MK-801 exposure of neurons experiencing normal spontaneous synaptic activity. After recording of whole-cell NMDAR currents, neurons were switched to current-clamp mode in normal  $Mg^{2+}$ -containing, TTX-free medium in the presence of MK-801 for the indicated times, n=3-6 per timepoint. This protocol is referred to as “activity block”. B) Comparison of the blockade of NMDARs by MK-801 under different conditions. “Quantal block” refers to blockade by MK-801 in zero  $Mg^{2+}$  + TTX, as described in (C). “Activity block” refers to blockade by MK-801 in normal  $Mg^{2+}$ -containing, TTX-free medium. “TTX” refers to blockade (or absence thereof) by MK-801 in normal  $Mg^{2+}$ -containing medium containing TTX. “Activity block+quantal block” refers to the application of the protocol described in (A) followed by the protocol described in (C). \* $p < 0.05$  (unpaired t-Test), n=7 (activity block), n=3 (TTX treatment), n=8 (quantal block), n=5 (activity block+quantal block). C) Loss of whole-cell current due to MK-801 exposure under quantal transmission plateaus after 10 min (n=3). Neurons were placed under voltage clamp and whole-cell NMDAR-mediated currents were measured. Neurons were then placed in  $Mg^{2+}$ -free external recording solution containing TTX and MK-801 for the indicated times, to allow open-channel blockade of synaptic NMDARs following their activation by quantal release of glutamate. D) Example of a whole-cell current trace before and after treatment described in (A). E) Confirmation that all synaptic NMDARs are blocked by the procedure described in (C). Waveforms representing an average of >200 mEPSCs before and after 10 min of MK-801 block of NMDARs activated by quantal transmitter release. Also for comparison is a waveform average of >200 mEPSCs recorded at the end of the experiment where all NMDARs were blocked by addition of a high concentration of agonist in the presence of MK-801. Note that the mEPSC timecourse in this instance is similar to that of mEPSCs recorded after 10 min “quantal block” protocol. F) Cell death due to 24 h  $H_2O_2$  (100  $\mu M$ ) insult in the face of the indicated treatments, applied 12 h before insult. \* $p < 0.05$  compared to control (n=3). G) Spontaneous EPSC frequency measured in the presence or absence of MK-801 (10  $\mu M$ ) to verify that it

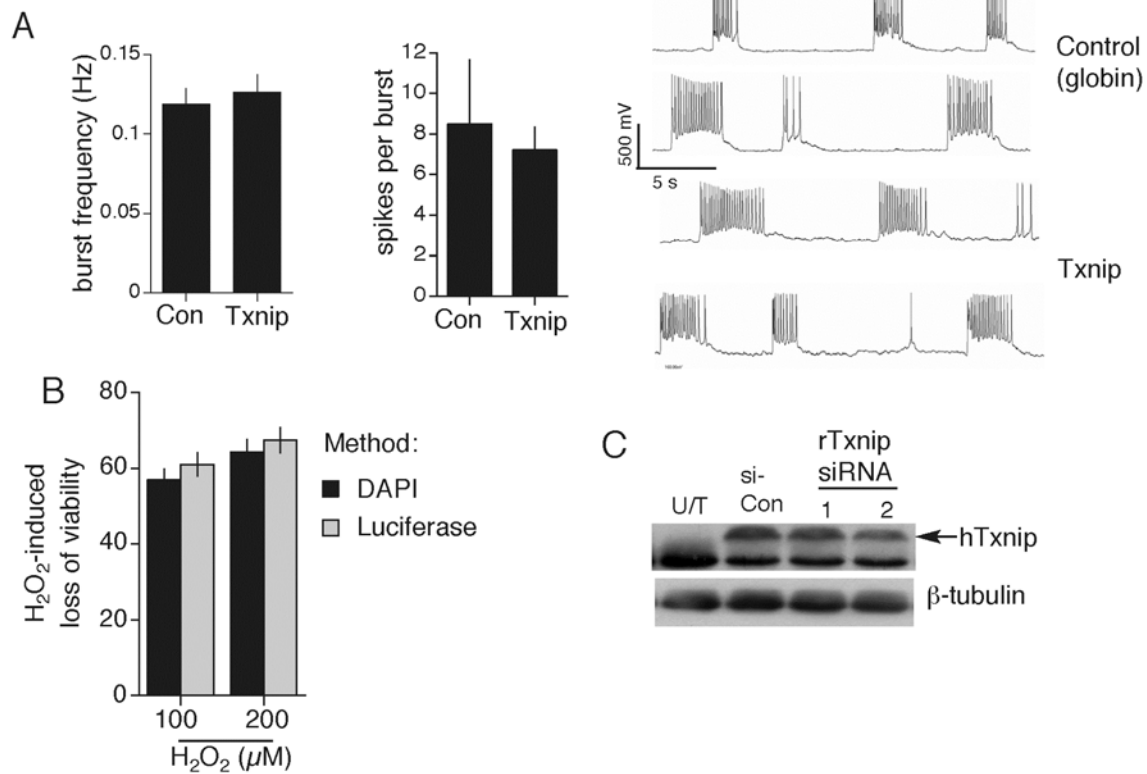
has no inherent effect on spontaneous electrical activity, emphasising that NMDAR activity and not electrical activity *per se*, is the key regulator of vulnerability to oxidative stress. H) Cell death due to 24 h H<sub>2</sub>O<sub>2</sub> (100 μM) insult in the face of the indicated treatments, applied 12 h before insult (n=3). BiC/4-AP stimulation is labelled as “BiC” in this and subsequent figures. I) Cell death due to 24 h H<sub>2</sub>O<sub>2</sub> (100 μM) insult in the face of the indicated treatments, applied 12 h before insult. Nifedipine was used at 5 μM. J) Cell death due to 24 h H<sub>2</sub>O<sub>2</sub> (100 μM) insult in the face of the indicated treatments, applied 12 h before insult. ω-conotoxin GVIA was used at 1 μM. K) Cell death due to 24 h H<sub>2</sub>O<sub>2</sub> (100 μM) treatment, in the presence ± Q-VD-Oph (50 μM) applied 1 h before H<sub>2</sub>O<sub>2</sub> treatment (n=4). L) Neurons treated with BiC/4-AP ± MK for 12 h prior to overnight treatment with H<sub>2</sub>O<sub>2</sub> (100 μM). Cell extracts were then assayed for caspase 9 and 3/7 activity (see methods) which were normalized to protein levels assayed by BCA assay \*p<0.05 (n=3).

Fig. S2



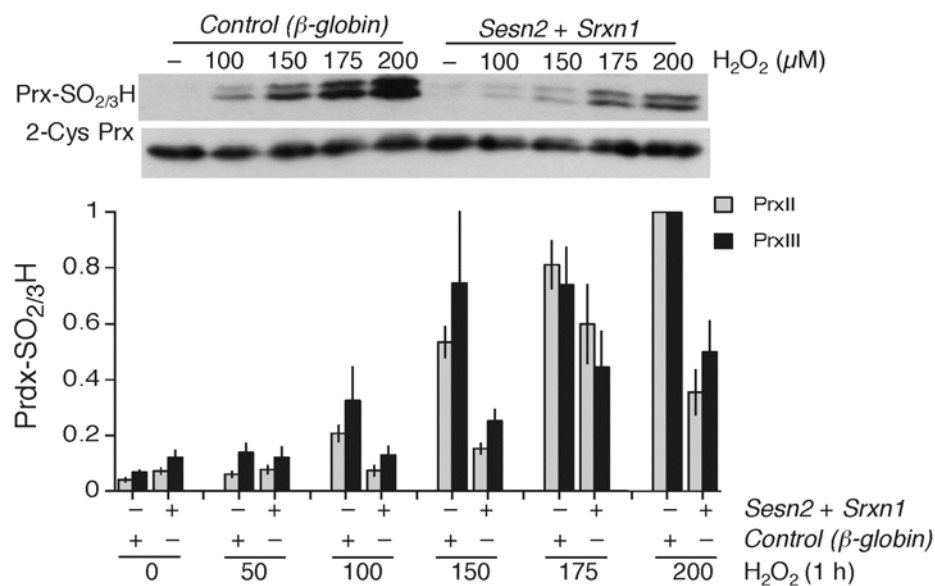
**Fig. S2.** A) PrxII, III and IV were ectopically expressed in cortical neurons, and protein extracts subjected to western analysis with a pan-2-Cys Prx antibody. A \* indicates which of the endogenous bands is enriched in each case. PrxIV expression enriches upper band by 10-fold, PrxIII enriches middle band by 2-fold, PrxII enriches bottom band by 2-fold. B) Western analysis of Prx overoxidation using an anti-PrxSO<sub>2/3</sub>H specific antibody. Analysis involved normalisation to appropriate Prx band intensity. \*p<0.05 compared to control, H<sub>2</sub>O<sub>2</sub>-treated neurons (n=4). C) Conventional RT-PCR illustrating time course of activity-dependent suppression, and activity blockade-dependent elevation, of *Txnip*, in comparison to *c-fos*. D) qPCR-based analysis of *Txnip* expression after 24 h treatment with the indicated drugs (MK-801 at 10 μM, TTX at 1 μM, n=4). E) Example pictures of immunofluorescent staining of cortical cultures with anti-NeuN (upper, to identify neurons), -GFAP (middle, to identify glial cells) and NeuN/GFAP-double-staining (lower). F) Example pictures of immunofluorescent staining of glial cultures with an anti-GFAP antibody.

Fig. S3



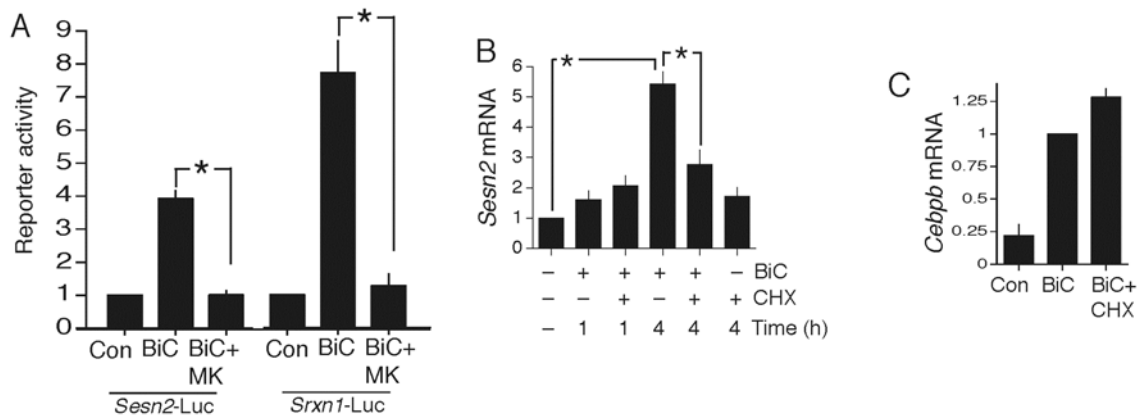
**Fig. S3.** A) BiC/4-AP-induced bursting was analyzed in neurons transfected with either Txnip- or globin-expression vectors, plus peGFP used to identify transfected cells. BiC/4-AP-induced bursting was recorded under current clamp and the burst frequency (left) and spikes/burst (middle) measured across 4 control and 4 Txnip expressing neurons. Example traces are shown on the right. B) Comparison of DAPI staining with constitutive luciferase expression as a metric of H<sub>2</sub>O<sub>2</sub>-induced cell death. Neurons were transfected with a constitutively active vector (SV40-Luc) and treated with H<sub>2</sub>O<sub>2</sub>, followed by luciferase activity measurement. In a parallel experiment, cell death was assessed conventionally by counting apoptotic nuclei stained with DAPI (n=3). C) Neurons were transfected with an expression vector for human Txnip plus control or one of two rat Txnip-directed siRNAs. Protein was harvested at 24 h and subjected to Western analysis for Txnip protein. Untransfected sample (U/T) shown for comparison.

Fig. S4



**Fig. S4.** HEK293 cells were transfected with expression vectors for Sesn2 and Srxn1, or control vector (globin). They were then treated with the indicated doses of H<sub>2</sub>O<sub>2</sub> and after 1 h harvested and subjected to western analysis of Prx-SO<sub>2/3</sub>H levels. Upper panel shows example western, lower panel shows analysis (n=3-5 westerns per timepoint).

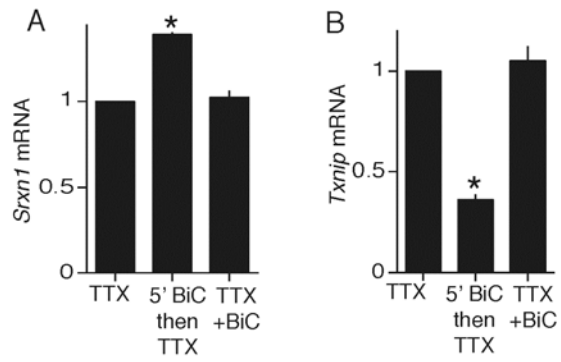
Fig. S5



**Fig. S5.** A) BiC/4-AP induces luciferase-based reporters of both *Sesn2* and *Srxn1* promoters in an MK-801-dependent manner. \* $p < 0.05$  ( $n = 5$ ). B) q-RT-PCR of *Sesn2* induction by synaptic activity at 4 h in the presence or absence of cycloheximide (10  $\mu\text{g/ml}$ ,  $n = 3$ ). C) q-RT-PCR of C/EBP $\beta$  induction by synaptic activity at 1 h in the presence or absence of cycloheximide ( $n = 3$ ).

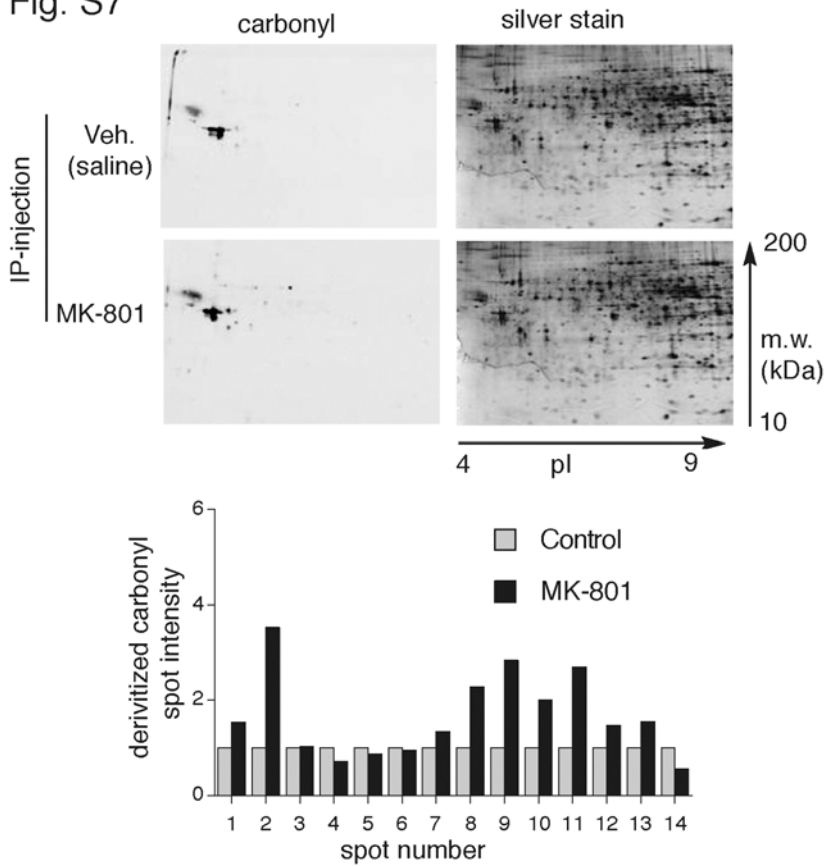


Fig. S6



**Fig. S6.** A) Analysis of *Srxn1* expression in neurons stimulated with BiC/4-AP for 5 min, after which activity was blocked by the addition of TTX. Expression was analyzed at 4 h and compared to TTX-treated neurons and to neurons treated with BiC/4-AP 2 min *after* TTX treatment. \* $p < 0.05$  (n=3). B) As for (A) but *Txnip* levels were analyzed at 1 h. \* $p < 0.05$  (n=3)

Fig. S7



**Fig. S7.** Analysis of carbonyl content of 2-D separated proteins from the cortices of adult mice subjected to IP-injection of MK-801. Carbonyl content (left) was detected using an antibody specific to the dinitrophenylhydrazone-derivatized residues. Blotted 2-DE gels were stained with silver staining to reveal protein spots (right). Below is a comparison of intensity of representative protein spots \* $p < 0.05$  ( $n=6$ ). Note that unlike P7 mice, none achieve significance, although a few spots approach significance.

## Supplemental Tables T1 and T2

Probe Set ID	Gene Title	Gene Symbol	RefSeq Transcript ID	Fold change (MK-801 <i>in vitro</i> )	Fold change (MK-801 <i>in vivo</i> )	Fold change (BiC/4-AP <i>in vitro</i> )
<b>Genes up-regulated by synaptic NMDAR activity</b>						
1422790_at	natriuretic peptide precursor type C	Nppc	NM_010933	-4.08	-1.75	2.57
1448285_at	regulator of G-protein signaling 4	Rgs4	NM_009062	-3.86	-3.20	2.33
1417065_at	early growth response 1	Egr1	NM_007913	-2.90	-2.15	2.31
1419248_at	regulator of G-protein signaling 2 adenylate cyclase activating	Rgs2	NM_009061	-2.51	-2.62	2.97
1423427_at	polypeptide 1 mitogen activated protein kinase	Adcyap1	NM_026249	-2.39	-2.63	4.40
1421340_at	kinase kinase 5	Map3k5	NM_009046 NM_009986	-2.13	-1.72	3.71
1415834_at	dual specificity phosphatase 6	Dusp6	/NM_198602	-2.13	-1.63	2.33
<b>Genes down-regulated by synaptic NMDAR activity</b>						
1415996_at	thioredoxin interacting protein	Txnip	NM_023719	5.02	2.62	-3.86

**Table T1.** List of genes whose expression is changed by BiC/4-AP treatment (in an MK-801 sensitive manner), and changed in the opposite direction by MK-801, both *in vitro* and *in vivo*.

	age	sex	case number	post-mortem interval	cause of death
<b>young</b>	25	F	1455	7	multiple injuries after vehicle accident
	26	M	a01-78	8	lung transplant, sepsis
	26	F	1489	16	suicide, drowning, history of depression
	35	M	1104	12	vehicle accident
	27	F	1614	18	gunshot wound to abdomen
	30	F	856	7	asthma
	37	F	a03-215	13	unknown
<b>old</b>	81	M	542	03:15	cardiac arrest
	82	M	86756591	04:30	unknown
	89	M	48046480	2	unknown
	82	F	963	8	dissecting aortic aneurysm
	83.7	M	885	2	urinary bladder rupture
	88	F	1037	20:30	lung cancer
	>90	M	911	07:24	Myocardial infarct
	95	M	1158	19	adenocarcinoma

**Table T2.** Information regarding the human samples analysed for Txnip expression