## **Supporting Information**

## Bacterial biosynthesis and maturation of the didemnin anticancer agents

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## **Supporting Information**

## **Supplementary Figures**



Fig. S1. Characterization of didemnin B. (a) MS<sup>n</sup> analysis.



Fig. S1. Characterization of didemnin B. (b) <sup>1</sup>H NMR spectrum.



Fig. S2. Characterization of nordidemnin B. (a) MS<sup>n</sup> analysis.



Fig. S2. Characterization of nordidemnin B. (b)  $^1\!H$  NMR spectrum.



Fig. S3. Bioinformatic analysis of uncharacterized *nrps1* gene cluster on plasmid 2 in the *T. mobilis* KA081020-065 genome. Domain notation: AL, acyl ligase; C, condensation domain; A, adenylation domain; T, thiolation domain; E, epimerization; TE, thioesterase. Substrate abbreviation: Dpg, dihydroxyphenylglycine.



NRPS or PKS related

ted F

Regulation, resistance related

Unknown function

4Kb

Gene	Size [aa]	Sequence similarity/organism	Proposed function	ldentity/ similarity	GenBank accession no.
orf1	261	Thioesterase, Actinomadura kijaniata	Thioesterase	41%, 54%	ACB46473
orf2	483	Membrane protease subunit stomatin/prohibitin-like protein, a- proteobacterium BAL199	Putative protease	42%, 59%	ZP_02189733
orf3	560	Cyclic peptide transporter, <i>Methylobacter tundripaludum</i> SV96	Transporter	48%, 68%	ZP_07653047
orf4	70	None	Hypothetical		
orf5	190	GTPase domain-containing protein, <i>Methylobacter tundripaludum</i> SV96	Hypothetical	39%, 57%	ZP_07653048
orf6	987	Hydrophobic/amphiphilic exporter-1, <i>Azospirillum</i> sp. B510	Resistance	38%, 56%	YP_003450508
orf7	377	Secretion protein, Azospirillum sp. B510	Resistance	28%, 47%	YP_003450507
didA	2123	OciA protein, <i>Planktothrix rubescens</i> NIVA-CYA 98	NRPS	29%, 40%	CAQ48254
didB	1796	Linear gramicidin synthetase subunit D, <i>Stigmatella aurantiaca</i> DW4/3-1	NPRS	36%, 48%	ZP_01459555
didC	1330	NRPS, Myxococcus xanthus DK 1622	NRPS	41%, 52%	YP_632257
didD	3853	Amino acid adenylation domain protein, <i>Streptomyces violaceusniger</i> Tu 4113	NRPS	39%, 50%	ZP_07603194
didE	1705	Amino acid adenylation domain protein, Acetivibrio cellulolyticus CD2	NRPS/PKS	36%, 53%	ZP_07325073
didF	1613	HctF, <i>Lyngbya majuscula</i>	NRPS	35%, 52%	AAY42398
didG	1413	NRPS/PKS, Amycolatopsis mediterranei U32	PKS	46%, 56%	YP_003765866
didH	1286	NRPS/PKS, Myxococcus xanthus DK 1622	NRPS	39%, 53%	YP_631961
didl	873	NRPS, Myxococcus xanthus DK 1622	NRPS	39%, 52%	YP_632257
didJ	2163	Amino acid adenylation domain protein, <i>Lyngbya majuscula</i> 3L	NRPS	36%, 53%	ZP_08431746

Fig. S4. Organization and deduced functions of the open reading frames within and flanking the didemnin biosynthetic gene cluster.

Gene	Size [aa]	Sequence similarity/organism	Proposed function	Identity/ similarity	GenBank accession no.
orf8	77	MbtH domain-containing protein, <i>Herpetosiphon aurantiacus</i> ATCC 23779	MbtH-like protein	80%, 89%	YP_001542806
orf9	68	Hypothetical protein, <i>Acidovorax</i> sp. JS42	Hypothetical	79%, 91%	YP_986866
orf10	45	None	Hypothetical		
orf11	60	None	Hypothetical		
orf12	190	Hypothetical protein, <i>Acidovorax</i> sp. JS42	Hypothetical	94%, 97%	YP_986861
orf13	75	Hypothetical protein, <i>Acidovorax</i> sp. JS42	Hypothetical	98%,100%	YP_004387524
orf14	324	CAAX amino terminal protease family, <i>Synechococcus</i> sp. PCC 7335	Putative protease	30%, 49%	ZP_05035401
orf15	398	Cyanate transport system protein, <i>Pseudomonas syringae</i> pv. syringae 642	Transport	40%, 52%	ZP_07265073
orf16	255	GntR family transcriptional regulator, <i>Chromobacterium violaceum</i> ATCC 12472	Regulation	39%, 55%	NP_903400

Fig. S4, continued. Organization and deduced functions of the open reading frames within and flanking the didemnin biosynthetic gene cluster.



Fig. S5. Characterization of didemnin B derivative [Hysp]didemnin B produced by *Tistrella mobilis* KA081020-065 due to putative NRPS substrate promiscuity in DidF A8 domain.  $MS^n$  analysis of [Hysp]didemnin B. Hysp,  $\alpha$ -( $\alpha$ -hydroxy sec-butylacetyl) propionic acid.



Fig. S6. Detection of didemnin precursors in the extract of *Tistrella mobilis* KA081020-065 by MALDI-TOF MS. a – Nordidemnin X ( $[M+H]^{\dagger}$ , obs 1651.8 Da, calc 1651.9 Da), b – Didemnin X ( $[M+H]^{\dagger}$ , obs 1665.8 Da, calc 1666.0), c – Nordidemnin Y ( $[M+H]^{\dagger}$ , obs 1779.7 Da, calc 1780.0 Da), d – Didemnin Y ( $[M+H]^{\dagger}$ , obs 1793.7 Da, calc 1794.0 Da).



Fig. S7. Characterization of didemnin precursors by MS<sup>n</sup>. (a) MS<sup>n</sup> analysis of didemnin X.



Fig. S7. Characterization of didemnin precursors by MS<sup>n</sup>. (b) MS<sup>n</sup> analysis of didemnin Y.



Fig. S7. Characterization of didemnin precursors by  $MS^n$ . (c)  $MS^n$  analysis of nordidemnin X.



Fig. S7. Characterization of didemnin precursors by MS<sup>n</sup>. (d) MS<sup>n</sup> analysis of nordidemnin Y.



Fig. S8. Didemnin precursor hydrolysis assay. MALDI-TOF MS analysis of didemnin X hydrolysis time course in presence of *Tistrella mobilis* secreted proteome ((a), 1d growth in liquid culture, >10kDa protein cutoff filter) and in presence of sterile GYP medium ((b), negative control).

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Fig. S8. Didemnin precursor hydrolysis assay. MALDI-TOF MS analysis of didemnin Y hydrolysis time course in presence of *Tistrella mobilis* secreted proteome ((c), 1d growth in liquid culture, >10kDa protein cutoff filter) and in presence of sterile GYP medium ((d), negative control).

d

Features	Chromosome	pTM1	pTM2	pTM3	pTM4
Topology	Circular	Circular	Circular	Circular	Circular
Genome size (bp)	3919492	692874	690188	1126962	83885
G+C content	68.15%	68.26%	67.61%	68.08%	67.26%
CDs number	3565	605	602	942	72
Coding density	89.08%	90.38%	88.92%	88.55%	85.06%
Average CDs length (bp)	979	1035	1019	1059	991
Assigned function	2852	432	501	726	60
Conserved hypothetical	472	101	71	165	9
Hypothetical	244	72	30	51	3
rRNA operons	2	0	0	1	1
tRNA operons	41	0	1	9	3

Table S1. General genome features of Tistrella mobilis KA081020-065