

Supporting Information

Methods and Materials

General

Chemicals were purchased from Sigma Aldrich (St. Louis, MO) except when indicated. *p*-Benzylisothiocyanato-desferrioxamine was obtained from Macrocyclics, Inc. (Dallas, Texas). Chemicals were used without further manipulations unless stated. Metal-free ultrapure water (>18.2 MΩ·cm, Milli-Q, Millipore, Billerica, MA) was obtained by soaking in Chelex 100 resin (Bio-Rad Laboratories, Hercules, CA) overnight at a ratio of 5g resin per 100 ml water as indicated in the manufacturer's instructions. For dose calibration measurements, a Capintec CRC-55tR (Capintec, Ramsey NJ) calibrated for Zr-89 was utilized (Calibration # 465). Quantification of radioactivity in counts-per-minute (cpm) was obtained through a Perkin Elmer Wizard² 2480 automatic gamma counter with an energy window of 800-1000 keV normalized for Zr-89 (909 keV). Radiochemical purity and yields were determined via a Bioscan AR-2000 radioTLC plate reader equipped with Winscan version 3.13 software. Silica-gel impregnated glass-fiber instant thin-layer chromatography paper (ITLC-SG, Varian Inc.) was used for ITLC analysis with 50 mM DTPA, pH 7 as the mobile phase. Vivaspin 500 centrifugal filters with a 10 kDa MWCO was obtained from GE Healthcare (Amersham, UK).

⁸⁹Zr Radiolabeling

The production of Zr-89 was made by a proton irradiation of a solid yttrium foil target on an EBCO TR19/9 variable beam energy cyclotron (EbcO Industries Inc., British Columbia, Canada) using methods published previously.⁽¹⁾ After end-of-bombardment, Zr-89 was isolated from the target by passing through a column of hydroxamate modified resin essentially capturing the isotope of interest. Elution was done by washing the column with 1 M oxalic acid resulting in a >99.99% radionuclidic and radiochemical purity and an effective specific activity of 195-497 MBq/μg (5.28-13.4 mCi/μg).^(1, 2) Typical radiolabeling conditions involve the following protocols. A solution of ⁸⁹Zr-oxalate (~1 mCi) was added into a 1.5 ml vial containing 100 μL metal-free water. The pH was adjusted to ~ 7.0-7.2 with 1 M Na₂CO₃. Effervescence was observed as the neutralization process resulted in CO₂ evolution. After the desired pH was obtained, 125-140 μg (Mb-DFO: 1.56-1.75 nmol; Cys-Db-DFO: 2.5 – 2.8 nmol) minibody/diabody-DFO was added. The reaction was incubated at room temperature with intermittent mild shaking. After 1-1.5 h, the reaction was quenched with approximately 50 μL of 50 mM DTPA (pH~7). Crude radiolabeling yields were determined to be >95% using iTLC with the Zr-89 labeled proteins remaining close to the origin (*R_f*=0.30) while the free Zr-89 is found near the solvent front (*R_f*=0.65). Purification of ⁸⁹Zr-protein was performed using a PD10 size exclusion column with saline as eluent and further concentrated with Vivaspin 500(MWCO: 10 kDa) centrifugal filter. The final radiochemical purity was >99% based on iTLC analysis.

SDS Gel Electrophoresis

20 μg of each sample was diluted in 20 μl of saline and mixed in a 1:1 ratio with Tris-Glycine SDS Sample Buffer (Invitrogen). After boiling at 95° C for 10 minutes, the samples were run on a 4-20% Tris-Glycine Gel (Invitrogen) with Tris-Glycine SDS Running Buffer (Invitrogen) at 150 V for 45 minutes. The gel was washed with 100 mL of ultrapure water for 15 minutes, then developed using Simple Blue Sage Stain (Invitrogen) for 1 hour and washed over night with 100 mL of ultrapure water (**SI Fig. 1**).

Supplemental Table 1. Biodistribution of ⁸⁹Zr-Mb administered via lateral tail vein in male athymic nude mice bearing PSMA(+) LNCaP prostate xenografts.

Tissue	1 h	4 h	12 h	24 h
	n=4	n=4	n=4	n=3
Blood	14.21 ± 2.52	6.64 ± 2.62	3.34 ± 0.71	1.99 ± 0.73
Tumor	2.29 ± 0.49	4.71 ± 0.98	6.12 ± 2.03	12.12 ± 3.61
Heart	4.50 ± 0.86	2.48 ± 0.78	2.13 ± 0.53	1.57 ± 0.45
Lungs	4.25 ± 1.76	2.36 ± 1.33	1.78 ± 0.80	1.98 ± 1.10
Liver	2.26 ± 0.72	1.47 ± 0.68	2.74 ± 0.40	5.06 ± 2.57
Spleen	1.52 ± 0.35	1.33 ± 0.66	1.47 ± 0.18	1.77 ± 0.96
Stomach	0.24 ± 0.10	0.17 ± 0.09	0.39 ± 0.26	0.73 ± 1.14
Sm. Intestines	1.39 ± 0.62	0.50 ± 0.23	0.48 ± 0.08	0.54 ± 0.21
L. Intestines	0.10 ± 0.03	0.78 ± 0.20	1.36 ± 0.62	0.53 ± 0.31
Kidney	6.98 ± 1.74	6.58 ± 1.54	6.29 ± 0.94	10.67 ± 4.42
Bone	3.78 ± 1.10	3.39 ± 0.59	2.77 ± 0.69	1.55 ± 0.17
Muscle	0.45 ± 0.10	0.39 ± 0.17	0.30 ± 0.04	0.43 ± 0.20
<i>Tumor/Blood</i>	<i>0.17 ± 0.06</i>	<i>0.81 ± 0.36</i>	<i>1.75 ± 0.33</i>	<i>5.93 ± 1.97</i>
<i>Tumor/Liver</i>	<i>1.05 ± 0.28</i>	<i>2.63 ± 0.09</i>	<i>1.88 ± 0.58</i>	<i>1.89 ± 0.20</i>
<i>Tumor/Kidneys</i>	<i>0.35 ± 0.13</i>	<i>0.65 ± 0.04</i>	<i>0.80 ± 0.35</i>	<i>0.90 ± 0.21</i>
<i>Tumor/Muscle</i>	<i>6.13 ± 0.53</i>	<i>11.35 ± 3.86</i>	<i>19.37 ± 5.48</i>	<i>26.81 ± 7.94</i>

Supplemental Table 2. Biodistribution of ⁸⁹Zr-Mb administered via lateral tail vein in male athymic nude mice bearing PSMA(-) PC3 prostate xenografts.

Tissue	1 h	4 h	12 h	24 h
	n=4	n=4	n=4	n=4
Blood	13.98 ± 1.46	6.94 ± 0.98	2.76 ± 0.44	1.14 ± 0.26
Tumor	2.28 ± 0.40	2.10 ± 0.34	1.67 ± 0.52	2.36 ± 0.31
Heart	3.95 ± 0.96	2.40 ± 0.24	1.66 ± 0.26	1.40 ± 0.43
Lungs	6.23 ± 2.22	1.72 ± 0.47	1.39 ± 0.60	1.32 ± 0.81
Liver	2.83 ± 1.16	1.78 ± 0.22	3.21 ± 1.31	3.07 ± 1.71
Spleen	2.24 ± 0.78	1.18 ± 0.24	1.69 ± 0.79	1.22 ± 0.27
Stomach	0.32 ± 0.05	0.47 ± 0.28	0.44 ± 0.14	0.08 ± 0.03
Sm. Intestines	1.60 ± 1.01	0.80 ± 0.43	0.39 ± 0.13	0.31 ± 0.14
L. Intestines	0.39 ± 0.31	0.99 ± 0.14	1.51 ± 1.58	0.24 ± 0.09
Kidney	8.18 ± 3.11	4.83 ± 0.96	3.59 ± 2.14	7.96 ± 4.46
Bone	2.81 ± 0.26	1.60 ± 0.39	1.45 ± 0.83	1.31 ± 0.63
Muscle	0.42 ± 0.12	0.21 ± 0.04	0.42 ± 0.29	0.13 ± 0.03
<i>Tumor/Blood</i>	<i>0.16 ± 0.01</i>	<i>0.30 ± 0.01</i>	<i>0.61 ± 0.20</i>	<i>2.13 ± 0.35</i>
<i>Tumor/Liver</i>	<i>0.81 ± 0.38</i>	<i>1.12 ± 0.09</i>	<i>0.54 ± 0.12</i>	<i>0.96 ± 0.46</i>
<i>Tumor/Kidneys</i>	<i>0.28 ± 0.13</i>	<i>0.44 ± 0.35</i>	<i>0.46 ± 0.24</i>	<i>0.36 ± 0.17</i>
<i>Tumor/Muscle</i>	<i>5.68 ± 1.46</i>	<i>11.86 ± 3.49</i>	<i>7.31 ± 3.24</i>	<i>18.10 ± 2.56</i>

Supplemental Table 3. Competitive binding with 200 µg and 500 µg non-radioactive Mb and huJ591.

Tissue	12 h p.i. (200 µg Mb)	12 h p.i. (500 µg Mb)	24 h p.i. (500 µg huJ591)
	n=6	n=4	n=4
Blood	4.08 ± 0.99	3.66 ± 0.68	2.76 ± 1.18
Tumor	3.73 ± 1.27	3.82 ± 0.70	4.98 ± 2.38
Heart	1.94 ± 0.55	2.06 ± 0.48	2.17 ± 0.58
Lungs	2.42 ± 0.98	1.22 ± 0.29	1.75 ± 0.93
Liver	4.24 ± 1.84	3.43 ± 1.21	3.49 ± 2.02
Spleen	1.60 ± 0.63	1.47 ± 0.65	2.08 ± 1.06
Stomach	0.34 ± 0.18	0.24 ± 0.12	0.32 ± 0.15
Sm. Intestines	0.52 ± 0.17	0.50 ± 0.23	0.45 ± 0.26
L. Intestines	0.47 ± 0.14	0.51 ± 0.23	0.95 ± 0.91
Kidney	9.60 ± 5.14	16.72 ± 6.14	8.61 ± 4.77
Bone	1.37 ± 0.81	2.98 ± 0.69	2.84 ± 1.46
Muscle	0.36 ± 0.23	0.36 ± 0.06	0.44 ± 0.14

Supplemental Table 4. Biodistribution of ⁸⁹Zr-Cys-Db administered via lateral tail vein in male athymic nude mice bearing PSMA(+) LNCaP prostate xenografts.

Tissue	1 h	4 h	12 h	24 h	12 h Block
	n=4	n=4	n=5	n=4	n=4
Blood	15.40 ± 4.23	4.44 ± 0.55	1.71 ± 0.43	0.59 ± 0.08	1.59 ± 0.18
Tumor	4.08 ± 1.00	6.91 ± 2.94	12.26 ± 2.54	6.53 ± 0.98	5.64 ± 1.75
Heart	5.50 ± 1.03	3.42 ± 0.95	2.61 ± 0.52	1.79 ± 0.06	2.81 ± 0.20
Lungs	7.35 ± 1.95	3.13 ± 1.17	1.97 ± 0.58	0.93 ± 0.24	1.23 ± 0.10
Liver	8.61 ± 0.80	5.40 ± 2.10	5.70 ± 2.79	5.54 ± 1.65	5.96 ± 1.69
Spleen	2.63 ± 0.40	2.54 ± 1.02	2.50 ± 1.57	1.75 ± 0.62	1.94 ± 0.83
Stomach	0.37 ± 0.03	0.59 ± 0.39	0.80 ± 0.38	0.59 ± 0.19	0.92 ± 0.40
Sm. Intestines	2.90 ± 1.06	1.26 ± 0.36	0.95 ± 0.38	0.74 ± 0.08	0.95 ± 0.27
L. Intestines	0.47 ± 0.22	1.57 ± 0.69	1.21 ± 0.40	0.77 ± 0.33	1.03 ± 0.33
Kidney	13.26 ± 5.92	16.86 ± 4.43	13.77 ± 1.94	15.47 ± 8.09	13.36 ± 1.79
Bone	4.48 ± 1.82	4.25 ± 1.54	3.69 ± 0.63	1.57 ± 0.52	5.09 ± 2.95
Muscle	0.72 ± 0.17	0.68 ± 0.18	0.47 ± 0.09	0.43 ± 0.12	0.44 ± 0.09
<i>Tumor/Blood</i>	<i>0.27 ± 0.03</i>	<i>1.24 ± 0.12</i>	<i>4.93 ± 0.89</i>	<i>11.19 ± 2.16</i>	
<i>Tumor/Liver</i>	<i>0.53 ± 0.08</i>	<i>1.25 ± 0.18</i>	<i>0.82 ± 0.18</i>	<i>1.23 ± 0.27</i>	
<i>Tumor/Kidneys</i>	<i>0.46 ± 0.17</i>	<i>0.50 ± 0.08</i>	<i>0.84 ± 0.23</i>	<i>0.53 ± 0.30</i>	
<i>Tumor/Muscle</i>	<i>4.90 ± 0.87</i>	<i>10.08 ± 2.31</i>	<i>15.55 ± 3.49</i>	<i>12.86 ± 1.72</i>	

Supplemental Table 5. Biodistribution of ⁸⁹Zr-Cys-Db administered via lateral tail vein in male athymic nude mice bearing PSMA(-) PC3 prostate xenografts.

Tissue	1 h	4 h	12 h	24 h
	n=5	n=4	n=5	n=4
Blood	10.39 ± 2.19	3.50 ± 0.41	0.96 ± 0.12	0.39 ± 0.12
Tumor	2.36 ± 0.48	3.64 ± 1.53	2.75 ± 0.51	3.44 ± 0.85
Heart	3.52 ± 0.98	2.98 ± 0.38	2.51 ± 0.26	2.10 ± 0.53
Lungs	5.14 ± 1.80	1.44 ± 0.11	1.15 ± 0.65	1.19 ± 0.47
Liver	5.05 ± 3.60	4.53 ± 1.32	5.39 ± 2.81	2.70 ± 0.37
Spleen	2.19 ± 1.10	1.72 ± 0.52	1.20 ± 0.36	0.93 ± 0.23
Stomach	0.52 ± 0.17	0.54 ± 0.24	0.77 ± 0.33	0.18 ± 0.09
Sm. Intestines	1.38 ± 0.50	0.94 ± 0.26	0.55 ± 0.23	0.40 ± 0.13
L. Intestines	1.15 ± 0.44	0.76 ± 0.13	0.92 ± 0.31	0.45 ± 0.09
Kidney	7.29 ± 2.61	12.74 ± 5.63	9.13 ± 2.68	9.10 ± 1.35
Bone	3.36 ± 0.80	3.60 ± 0.51	2.51 ± 1.70	1.81 ± 0.66
Muscle	0.82 ± 0.51	0.73 ± 0.25	0.54 ± 0.24	0.29 ± 0.05
<i>Tumor/Blood</i>	<i>0.24 ± 0.08</i>	<i>1.01 ± 0.34</i>	<i>2.87 ± 0.43</i>	<i>9.18 ± 1.76</i>
<i>Tumor/Liver</i>	<i>0.67 ± 0.44</i>	<i>0.89 ± 0.53</i>	<i>0.63 ± 0.31</i>	<i>1.06 ± 0.52</i>
<i>Tumor/Kidneys</i>	<i>0.36 ± 0.16</i>	<i>0.34 ± 0.21</i>	<i>0.32 ± 0.10</i>	<i>0.39 ± 0.15</i>
<i>Tumor/Muscle</i>	<i>3.62 ± 1.77</i>	<i>5.28 ± 2.19</i>	<i>5.68 ± 1.86</i>	<i>12.53 ± 5.08</i>

SI Table 6. Tumor uptake expressed as %ID/g (mean \pm S.D.) of all three radiotracers obtained from VOIs.

Time, h	$^{89}\text{Zr-Mb}$	$^{89}\text{Zr-Cys-Db}$	$^{89}\text{Zr-huJ591}$
1	3.23 \pm 0.96	5.58 \pm 1.09	2.94 \pm 0.66
4	6.16 \pm 1.43	8.19 \pm 1.41	10.21 \pm 1.95
12	6.85 \pm 0.87	9.84 \pm 2.54	15.84 \pm 1.79
24	7.93 \pm 0.67	9.17 \pm 1.48	26.57 \pm 3.50

SI Table 7. Blood clearance of all three radiotracers obtained from heart VOIs expressed as %ID/g (mean \pm S.D.).

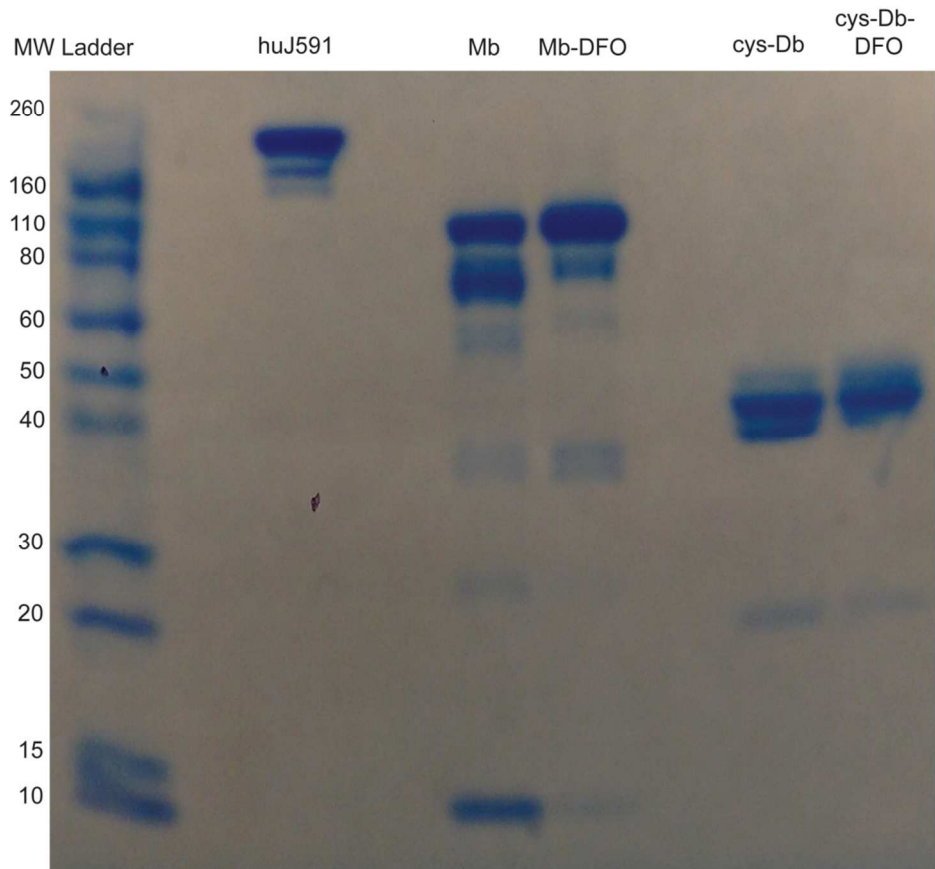
Time, h	⁸⁹Zr-Mb	⁸⁹Zr-Cys-Db	⁸⁹Zr-huJ591
1	20.86 \pm 3.39	13.19 \pm 2.70	25.49 \pm 2.11
4	13.90 \pm 3.06	5.90 \pm 1.10	21.66 \pm 1.91
12	5.06 \pm 2.84	2.74 \pm 0.61	16.36 \pm 1.57
24	3.26 \pm 1.20	2.00 \pm 0.18	12.13 \pm 1.26

SI Table 8. Comparison of tumor-to-muscle ratios of all radiotracers.

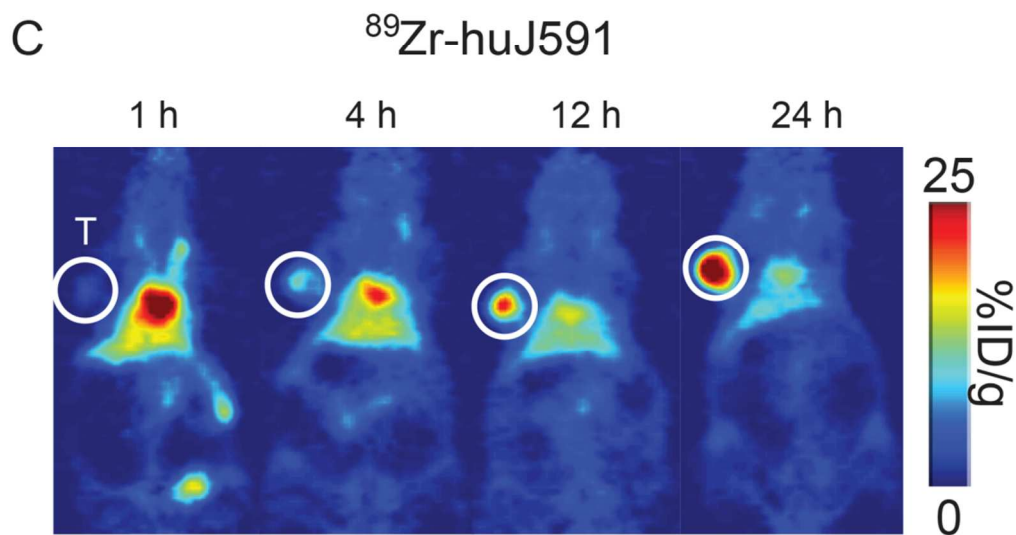
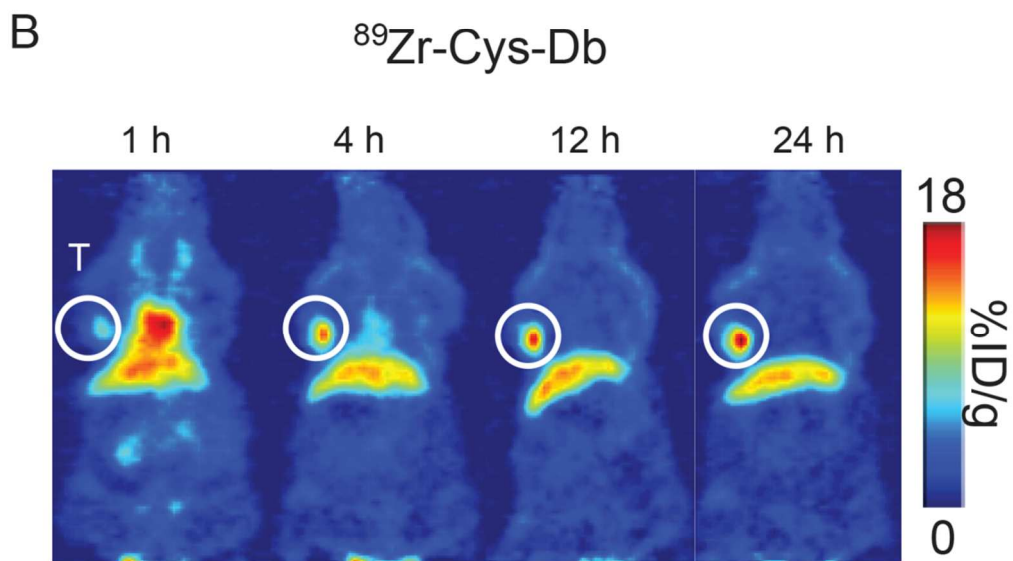
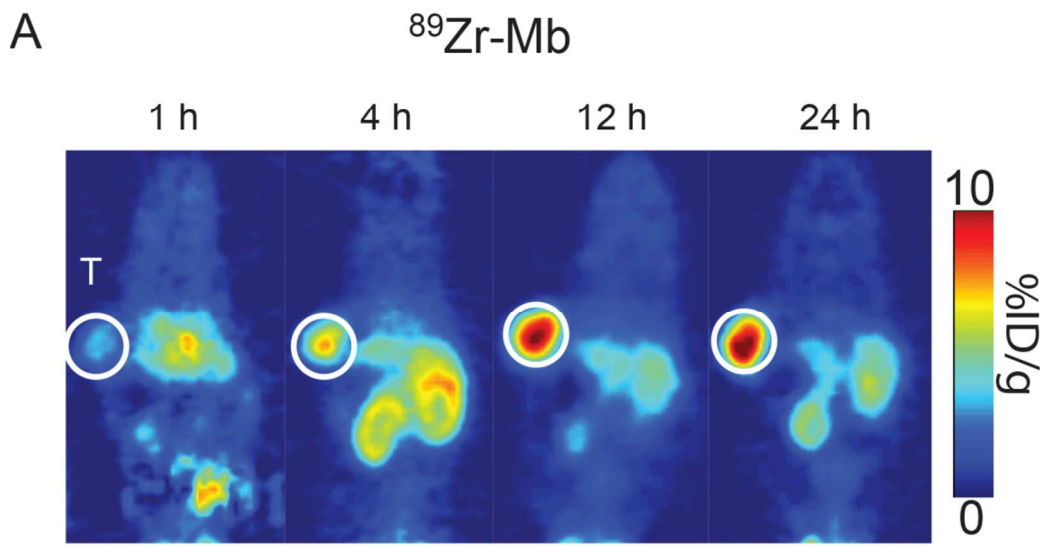
Time, h	⁸⁹Zr-Mb	⁸⁹Zr-Cys-Db	⁸⁹Zr-huJ591
1	2.53 ± 0.34	3.48 ± 0.48	2.92 ± 0.21
4	4.90 ± 0.96	5.17 ± 1.09	6.69 ± 1.79
12	10.04 ± 3.70	6.09 ± 0.85	7.87 ± 1.63
24	9.59 ± 3.68	6.89 ± 1.15	11.02 ± 2.30

SI Table 9. Comparison of tumor-to-blood (heart) ratios of all radiotracers.

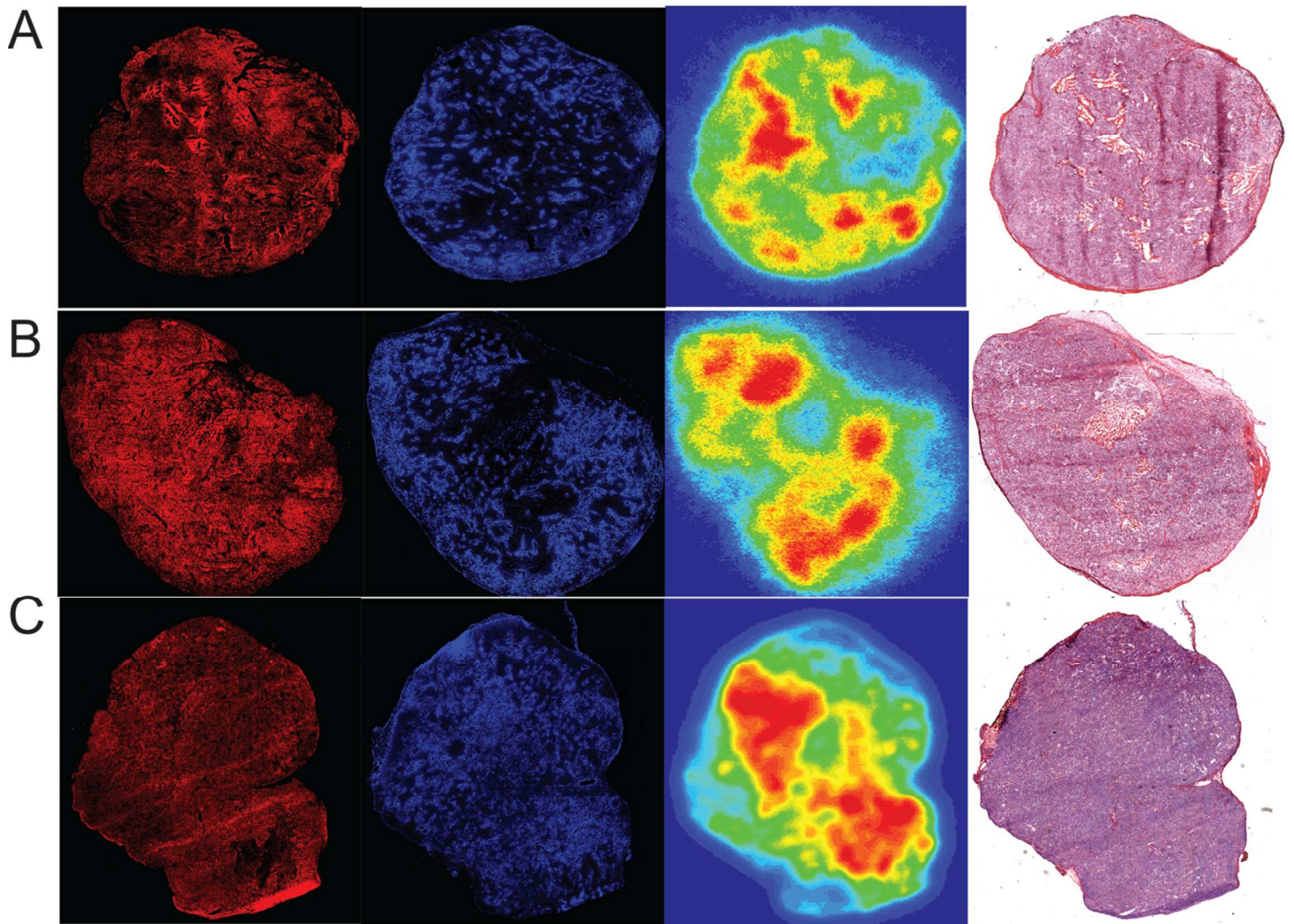
Time, h	⁸⁹Zr-Mb	⁸⁹Zr-Cys-Db	⁸⁹Zr-huJ591
1	0.15 ± 0.03	0.43 ± 0.09	0.12 ± 0.02
4	0.41 ± 0.02	1.40 ± 0.22	0.47 ± 0.09
12	1.67 ± 0.81	3.58 ± 0.19	0.98 ± 0.19
24	2.65 ± 0.90	4.91 ± 0.26	2.22 ± 0.46



SI Figure 1. SDS PAGE gel electrophoresis of huJ591, DFO-derivatized and unmodified Mb and Cys-Db.



SI Figure 2. Planar sections of serial PET images obtained with $^{89}\text{Zr-Mb}$ (A), $^{89}\text{Zr-Cys-Db}$ (B) and $^{89}\text{Zr-huJ591}$ from 1-24 h post-injection.



SI Figure 3. Autoradiography and Histology. Hoechst 33342 (blue), PSMA staining (red), autoradiographs and H&E stain of (A) ^{89}Zr -Mb and (B) ^{89}Zr -Cys-Db and (C) ^{89}Zr -huJ591.

References:

1. Holland JP, Sheh Y, Lewis JS. Standardized methods for the production of high specific-activity zirconium-89. *Nucl Med Biol.* Oct 2009;36(7):729-739.
2. Verel I, Visser GW, Boellaard R, Stigter-van Walsum M, Snow GB, van Dongen GA. ^{89}Zr immuno-PET: comprehensive procedures for the production of ^{89}Zr -labeled monoclonal antibodies. *J Nucl Med.* Aug 2003;44(8):1271-1281.