Couturier et al., http://www.jcb.org/cgi/content/full/jcb.201407071/DC1

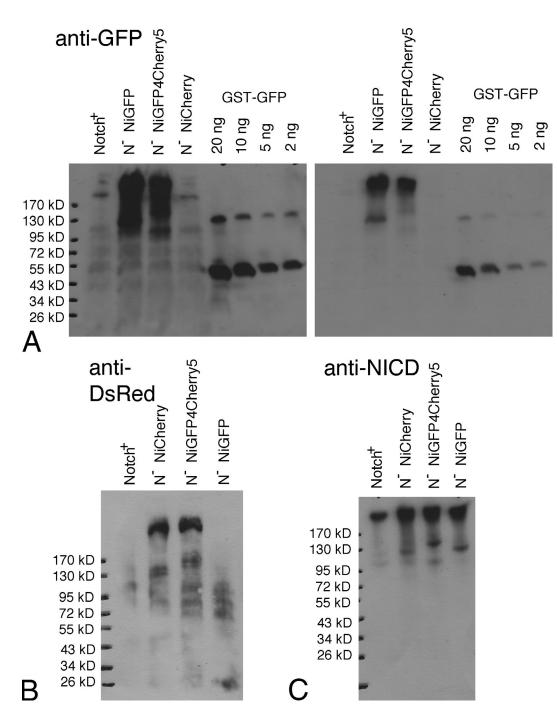
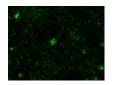


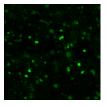
Figure S1. Western blot analysis of tagged Notch. (A–C) Anti-GFP (A), anti-DsRed (B; recognizing Cherry), and anti-NICD (C) Western blot analysis of total wing imaginal disc extracts. (A) A purified GST-GFP protein was loaded to evaluate the amount of GFP-tagged Notch in these extracts. (A) Anti-GFP specifically recognized full-length NiGFP and NiGFP4Cherry5 as well as  $\sim$ 140 (NiGFP]- and  $\sim$ 160 (NiGFP4Cherry5)-kD molecular species . These likely correspond to S1-cleaved receptors with the increase in size being attributable to the fluorescent protein tags. Note the absence of signal at the expected size for GFP. (B) Sinilarly, anti-DsRed specifically recognized full-length NiCherryP and NiGFP4Cherry5 and Smaller  $\sim$ 140/ $\sim$ 160-kD species. No proteo-lytically resistant GFP and/or Cherry molecules or other unexpected proteolytic fragments were detected (A and B), even on overexposed autoradiograms (A). Anti-NICD recognized full-length Notch, NiGFP, NiCherry, and NiGFP4Cherry5 as well as a  $\sim$ 140 (NiGFP and NiCherry]- and  $\sim$ 160 (NiGFP and NiCherry]- and  $\sim$ 160 (NiGFP Acherry5)-kD molecular species. No technologians (A). Anti-NICD recognized full-length Notch, NiGFP, NiCherry, and NiGFP4Cherry5 as well as a  $\sim$ 140 (NiGFP and NiCherry]- and  $\sim$ 160 (NiGFP 4Cherry5)-kD molecular species. N, Notch.

## Table S1. Genotypes used in this paper

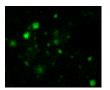
| Genotypes   | Figures                                 |
|---|---|
| N <sup>55e11</sup> w <sup>1118</sup> /Y; M[3×P3-RFP.attP.w <sup>+</sup> .NiGFP]51D/+  | Fig. 1, B and B'                        |
| N <sup>55</sup> e <sup>11</sup> w <sup>1118</sup> /Y; M[3×P3-RFP.attP.w <sup>+</sup> .NiCherry]51D/+  | Fig. 1, C and C'                        |
| N <sup>55e11</sup> w <sup>1118</sup> /Y; M[3×P3-RFP.attP.w <sup>+</sup> .NiGFP]51D/M[3×P3-RFP.attP.w <sup>+</sup> .NiCherry]51D   | Fig. 1, D and D'                        |
| N <sup>55e11</sup> w <sup>1118</sup> /Y; ; PB[y*.attP.w⁺.NiGFP4Cherry5]68D/+  | Fig. 1, E–F' and Fig. 3,<br>K and L     |
| 2[pRab-5-Cherry-Rab5] w <sup>1118</sup> /N <sup>55e11</sup> w <sup>1118</sup> P[neur-Histone2B-RFP]619 ; M[3×P3-RFP.attP.w⁺.NiGFP]51D/+   | Fig. 3, B and B'                        |
| v <sup>55e11</sup> w <sup>1118</sup> P[neur-Histone2B-RFP]619/Y; M[3×P3-RFP.attP.w <sup>+</sup> .NiCherry]51D/GFP-Rab5[KI]  | Fig. 3 C                                |
| V <sup>55e11</sup> w <sup>1118</sup> P[Ubx-flp] P[neur-Histone2B-RFP]619/Y; M[3×P3-RFP.attP.w <sup>+</sup> .NiCherry]51D/+; P[pTub-YFP-Rab11]/+   | Fig. 3 D                                |
| √ <sup>55e11</sup> w <sup>1118</sup> P[neur-Histone2B-RFP]619/Y ; M[3×P3-RFP.attP.w <sup>+</sup> .NiCherry]51D/+; P[pTub-GFP-Lamp1]/+   | Fig. 3 E                                |
| N <sup>55e11</sup> w <sup>1118</sup> P[Ubx-flp] P[neur-Histone2B-RFP]619/Y; M[3×P3-RFP.attP.w <sup>+</sup> .NiCherry]51D/+; P[pTub-YFP-Rab7]/+  | Fig. 3, F and G                         |
| √ <sup>55e11</sup> w <sup>1118</sup> /Y; ap-Gal4 P[pTub-Gal80 <sup>ts</sup> ]/+; PB[y <sup>+</sup> .attP.w <sup>+</sup> .NiGFP4Cherry5]68D/+  | Fig. 3 H                                |
| v <sup>1118</sup> /Y; ap-Gal4/+; PB[y <sup>+</sup> .attP.w <sup>+</sup> .NiGFP4Cherry5]68D/P{TRiP.HMS01287}attP2 (rbcn3A <sup>RNAi</sup> )  | Fig. 3 I                                |
| N <sup>55e11</sup> w <sup>1118</sup> P[Ubx-flp] P[neur-Histone2B-RFP]619/Y; P[neo, ry FRT40A] lgd <sup>47</sup> /P[neo, ry FRT40A] P[pTub-nlsRFP]; PB[y <sup>+</sup> .<br>attP.w <sup>+</sup> . NiGFP4Cherry5]68D/+ | Fig. 3, M and M'                        |
| CadGFP (KI)/CadCherry (KI)  | Fig. 5, A–A″                            |
| v <sup>1118</sup> /Y; ; P[y <sup>+</sup> .attP.w <sup>+</sup> .SpdoCherry2GFP3]68A4 spdo <sup>ZZ27</sup> /spdo <sup>G104</sup>  | Fig. 5 C                                |
| v <sup>1118</sup> /Y; ; P[y <sup>+</sup> .attP.w <sup>+</sup> .SpdoCherryGFP]99F8   | Fig. 5, D–D"; Fig. 6 D;<br>and Fig. 7 E |
| N <sup>55e11</sup> w <sup>1118</sup> /Y; P[pneur-nlsFP670]/+ ; PB[y⁺.attP.w⁺. NiGFP4Cherry5]68D/+   | Fig. 6 and Fig. 7, A–C"                 |
| P[Ubx-flp]/Y; P[neo, ry FRT40A] numb <sup>15</sup> /P[neo, ry FRT40A] P[pTub-nlsGFP]; P[y <sup>+</sup> .attP.w <sup>+</sup> .SpdoCherryGFP]99F8/+   | Fig. 7 F                                |



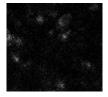
Video 1. Localization of NiGFP and Cherry-Rab5 in notum epithelial cells. Live imaging of GFP-tagged Notch (green) and Cherry-tagged Rab5 (red) in an N<sup>55e11</sup> H2B-RFP/Cherry-Rab5; NiGFP/+ pupa at 16 h APF showing a subapical confocal section. A large NiGFP-positive, Cherry-Rab5 endosome is indicated with an arrow. Images were analyzed by time-lapse confocal microscopy using a laser-scanning confocal microscope (LSM 580; Carl Zeiss). Time is in minutes and seconds.



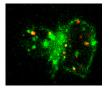
Video 2. Localization of NiGFP and Cherry-Rab4 in notum epithelial cells. Live imaging of GFP-tagged Notch (green) and Cherry-tagged Rab4 (red) in an N<sup>55e11</sup>/Y; Cherry-Rab4; NiGFP/+ pupa at 16 h APF. A subapical confocal section is shown. Images were analyzed by time-lapse confocal microscopy using a laser-scanning confocal microscope (LSM 580; Carl Zeiss). Time is in minutes and seconds.



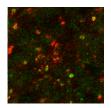
Video 3. **Localization of NiCherry and GFP-Rab5 in notum epithelial cells.** Live imaging of Cherry-tagged Notch (red) and GFP-tagged Rab5 (green) in an N<sup>55e11</sup> H2B-RFP/Y; NiCherry/GFP-Rab5 pupa at 16 h APF. A subapical confocal section is shown. Images were analyzed by time-lapse confocal microscopy using a laser-scanning confocal microscope (LSM 580; Carl Zeiss). The arrow points to a Rab5-positive endosome that contains NiCherry. Time is in minutes and seconds.



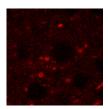
Video 4. **Localization of NiCherry and YFP-Rab7 in notum epithelial cells.** Live imaging of Cherry-tagged Notch (red) and YFP-tagged Rab7 (green) in an N<sup>55e11</sup> H2B-RFP/Y; NiCherry/YFP-Rab7 pupa at 16 h APF. A basal confocal section is shown. Images were analyzed by time-lapse confocal microscopy using a laser-scanning confocal microscope (LSM 580; Carl Zeiss). Time is in minutes and seconds.



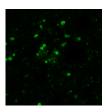
Video 5. Localization of SpdoCherryGFP in plla/pllb cells. Live imaging of SpdoCherryGFP (red and green) in a SpdoCherryGFP/+ pupa at 16 h APF. An endosomal fusion event seen in pllb is highlighted using a 4× magnification. plla/pllb cell pairs were imaged at *t* = 20 min after mitosis. Images were analyzed by time-lapse confocal microscopy using a laser-scanning confocal microscope (LSM 580; Carl Zeiss). Time is in minutes and seconds.



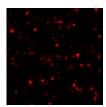
Video 6. Localization of NiGFP and Cherry-Rab5 in plla/pllb cells. Live imaging of GFP-tagged Notch (green) and Cherry-tagged Rab5 (red) in an  $N^{55e11}$  H2B-RFP/Cherry-Rab5; NiGFP/+ pupa at 16 h APF. The plla and pllb cells were identified on apical (weak NiGFP signal; left image) and basal (H2B-RFP–positive nuclei; central image). NiGFP-positive Cherry-Rab5 endosomes are indicated with a green arrow in pllb at t = 20 min after mitosis. Images were analyzed by time-lapse confocal microscopy using a laser-scanning confocal microscope (LSM 580; Carl Zeiss). Time is in minutes and seconds.



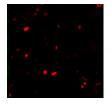
Video 7. Localization of SpdoGFP1 and Cherry-Rab5 in plla/pllb cells. Live imaging of GFP-tagged Spdo (green) and Cherrytagged Rab5 (red) in a *Cherry-Rab5/+; SpdoGFP1/+* pupa at 16 h APF. SpdoGFP1 colocalized with Cherry-Rab5 in pllb 20 min after mitosis (Spdo was used to identify plla/pllb cells). A subapical confocal section is shown. Images were analyzed by time-lapse confocal microscopy using a laser-scanning confocal microscope (LSM 580; Carl Zeiss). Time is in minutes and seconds.



Video 8. Localization of NiCherry and GFP-Rab5 in plla/pllb cells. Live imaging of Cherry-tagged Notch (red) and GFP-tagged Rab5 (green) in an  $N^{55e11}$  H2B-RFP/Y; NiCherry/GFP-Rab5 pupa at 16 h APF. The plla and pllb cells were identified using H2B-RFP (left). A subapical confocal section is shown in the video. NiCherry did not colocalize with GFP-Rab5 in pllb at t = 20 min after mitosis. Images were analyzed by time-lapse confocal microscopy using a laser-scanning confocal microscope (LSM 580; Carl Zeiss). Time is in minutes and seconds.



Video 9. Localization of NiCherry and SpdoGFP in pllb. Live imaging of Cherry-tagged Notch (red) and GFP-tagged Spdo (green) in an  $N^{55e11}$  H2B-RFP/+; NiCherry/+; SpdoGFP1/+ pupa at 16 h APF. The plla and pllb cells were identified using H2B-RFP and SpdoGFP1. A subapical confocal section is shown in the video. NiCherry did not colocalize with SpdoGFP1 in pllb at t = 20 min after mitosis. Images were analyzed by time-lapse confocal microscopy using a laser-scanning confocal microscope (LSM 580; Carl Zeiss). Time is in minutes and seconds.



Video 10. Localization of NiCherry and NumbGFP in pllb. Live imaging of Cherry-tagged Notch (red) and GFP-tagged Numb (green) in an NiCherry/+; NumbGFP/+ pupa at 16 h APF. The plla and pllb cells were identified using the unequal segregation of Numb at mitosis. A subapical confocal section is shown in the video. NiCherry did not colocalize with NumbGFP in pllb at t = 20 min after mitosis. Images were analyzed by time-lapse confocal microscopy using a laser-scanning confocal microscope (LSM 580; Carl Zeiss). Time is in minutes and seconds.