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Supplemental Information

Cancer Cells Upregulate NRF2

Signaling to Adapt to Autophagy Inhibition

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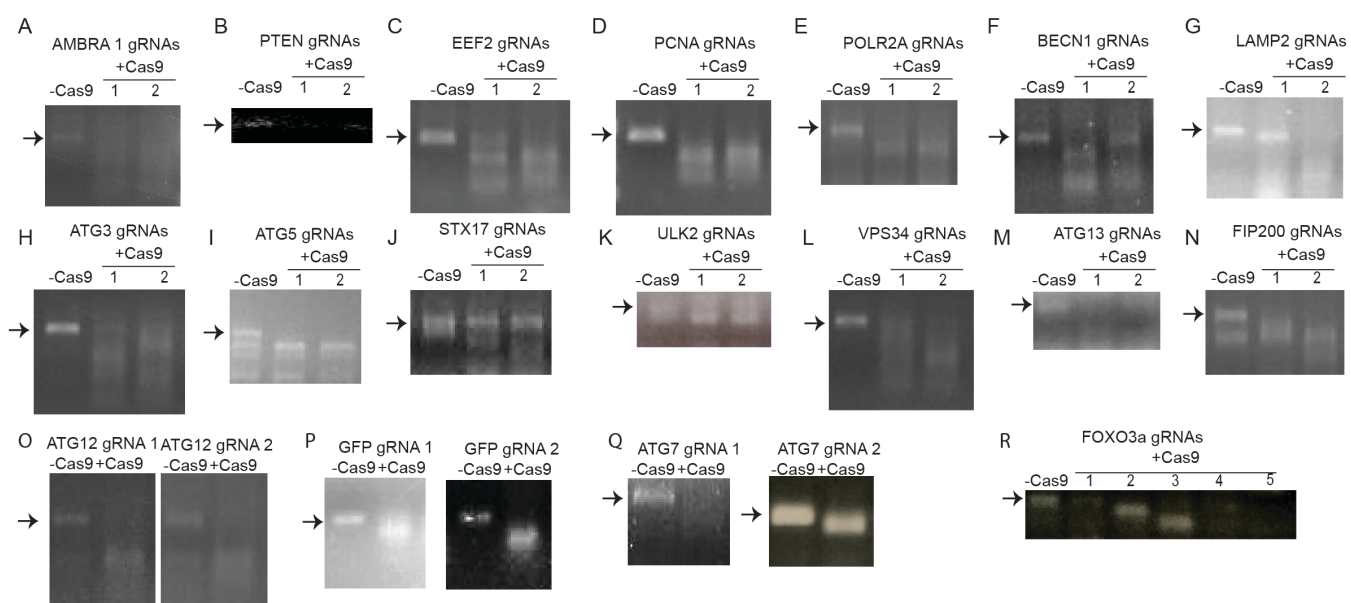


Figure S1, Related to Figure 1: Verification of gRNAs. Agarose gels showing cut DNA before and after In-vitro Cas9 assay for all the indicated gRNAs used in Figure 1 and 2. Black arrows indicate the correct PCR product running at the expected size.

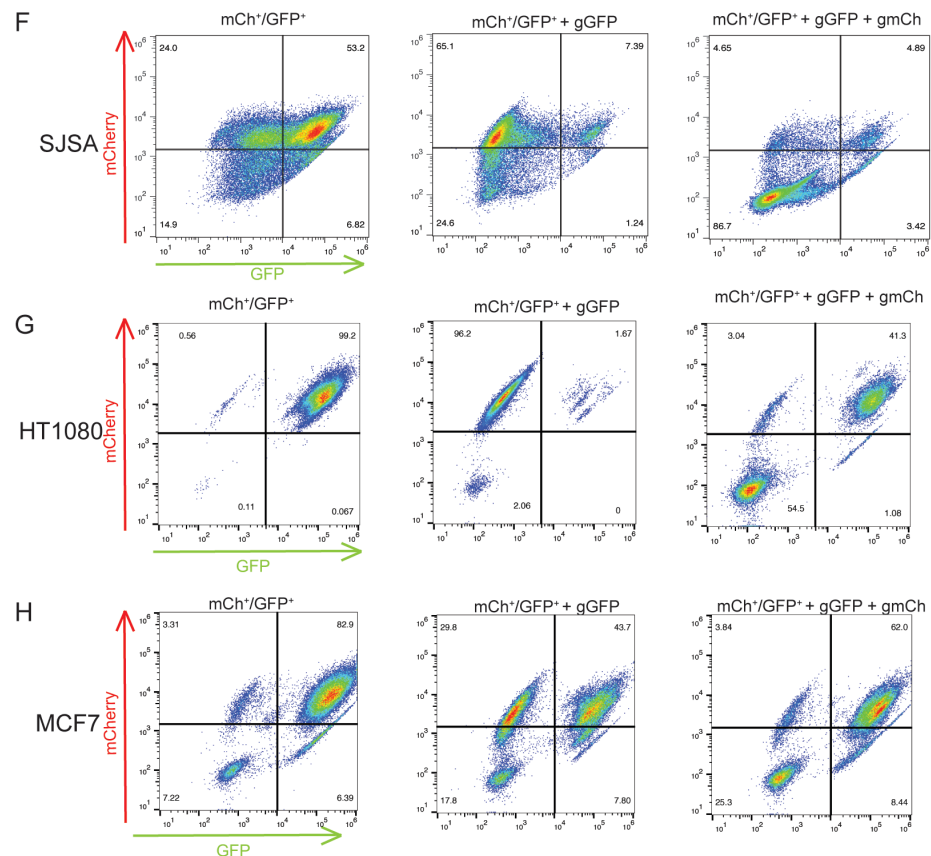
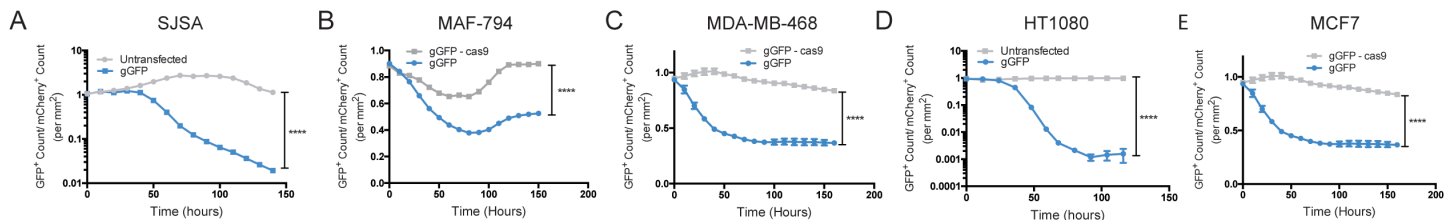
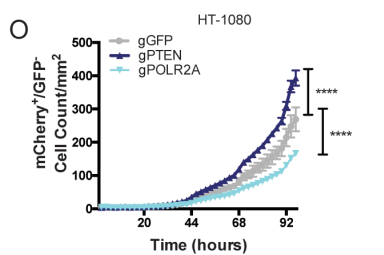
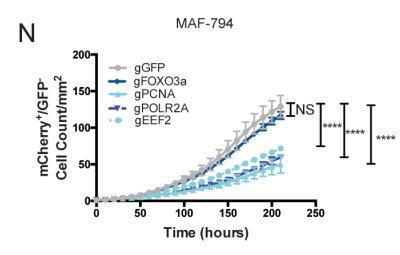
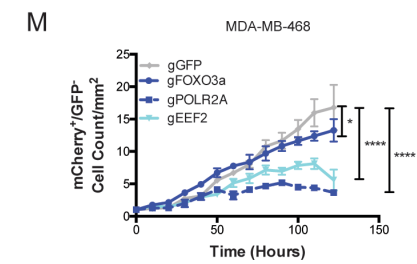
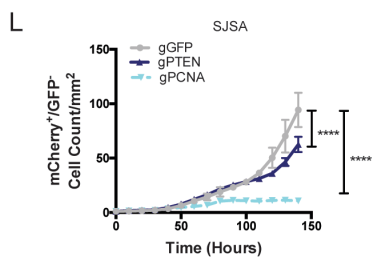
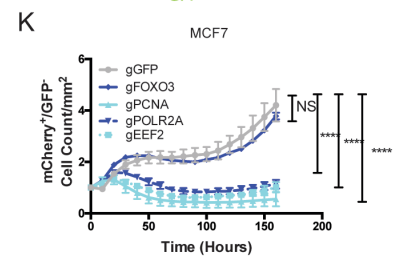
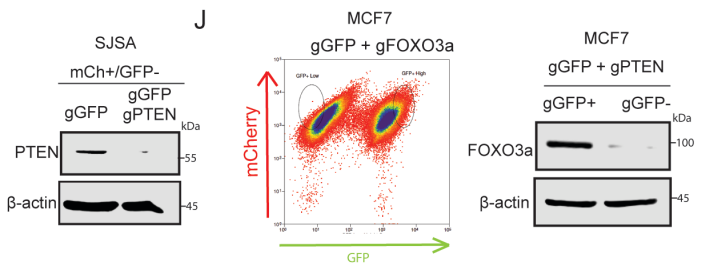
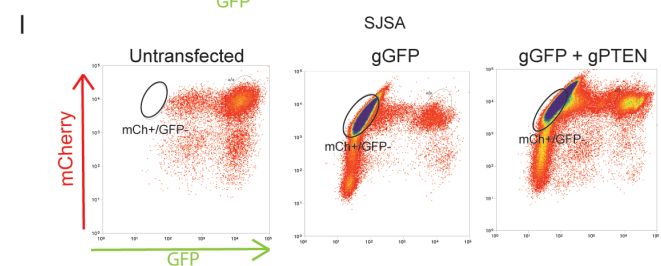


Figure S2, Related to Figure 1: Design of a quantitative live-cell imaging CRISPR-RNP assay to identify essential genes in multiple cell lines. (A-E) Incucyte quantification of GFP+ cell count normalized to mCherry+ cell count in mCherry+/GFP+ indicated cell lines after transfection with gRNAs targeting GFP. Data represented as mean±SEM for technical replicates (N of 2-3) and are representative of 2-3 individual experiments. Statistical analysis: 2-way ANOVA. (F-H) Flow cytometry was performed in mCherry+/GFP+ cells before and after transfection with gGFP or gGFP and gmCherry. The data are representative of multiple experiments (N of 2-3). (I-J) Western blots ran on the GFP- sorted populations from mCherry+/GFP+ cells subject to RNP transfections with gRNAs targeting GFP and the indicated genes. (K-O) Incucyte quantification of mCherry+/GFP- cell count/mm² after transfection with RNPs targeting GFP and the indicated genes. The data are represented as mean±SEM for technical replicates (N of 2-3) and the graphs shown are representative of 2-3 individual experiments. Statistical analysis: 2-way ANOVA and the significance at the last time point is shown. *p<0.05, **p<0.01, ***p<0.001 **** p<0.0001.



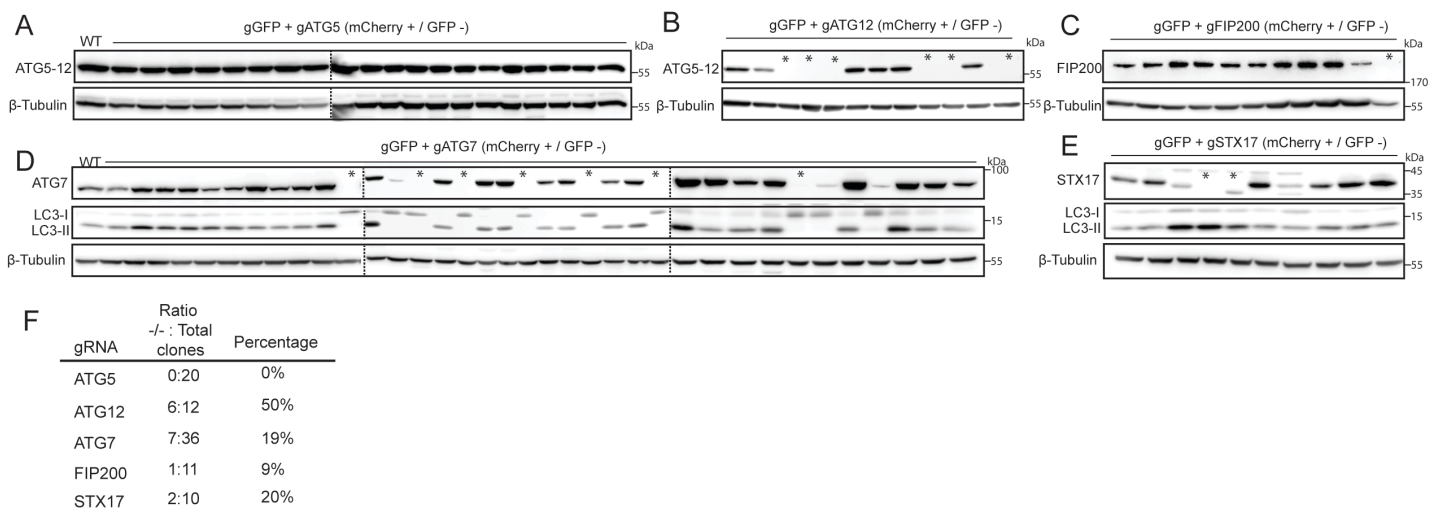
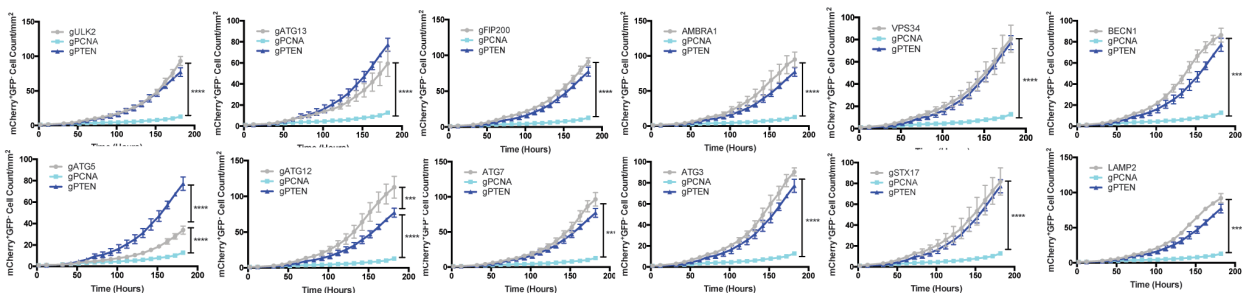


Figure S3, Related to Figure 2: Verification of gene KO in H292 cells. (A-E) Western blots ran on clones generated from the GFP- sorted populations from mCherry+/GFP+ cells subject to RNP transfections with gRNAs targeting GFP and the indicated genes. * represent clones with complete KO. (F) Calculated percent of clones with complete loss of protein expression based on western blots from A-E. Lorem ipsum

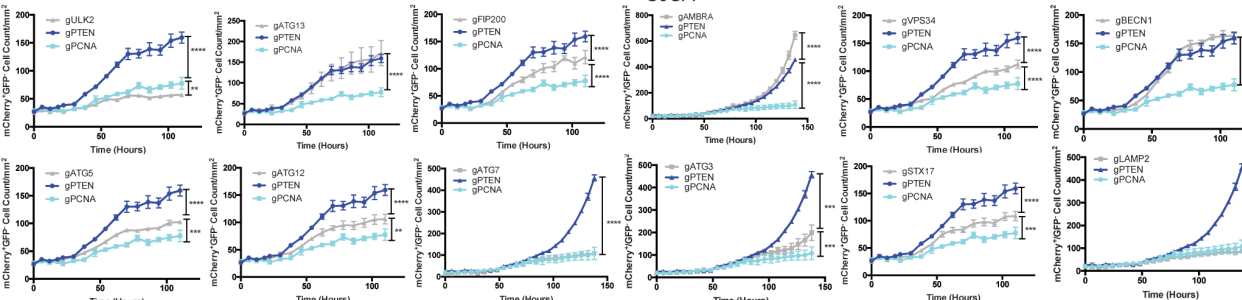
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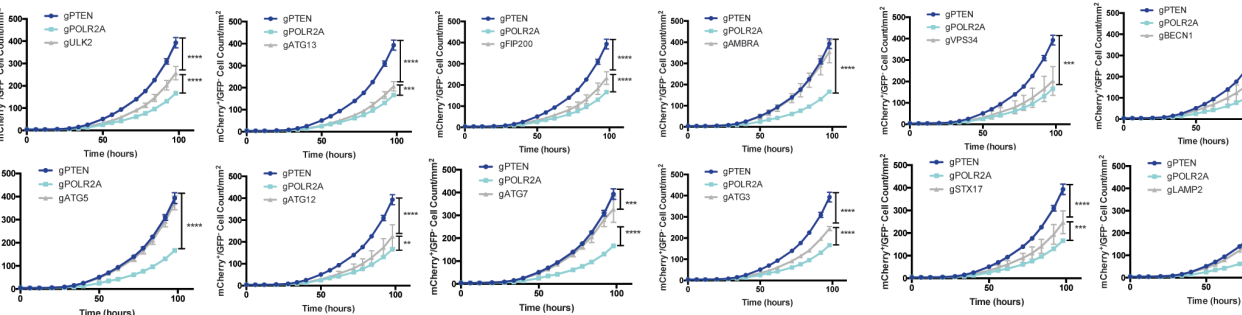
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SJSA



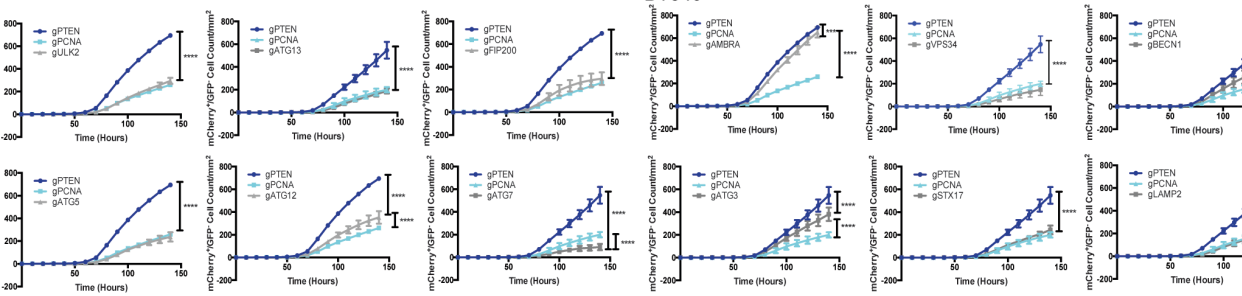
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HT1080



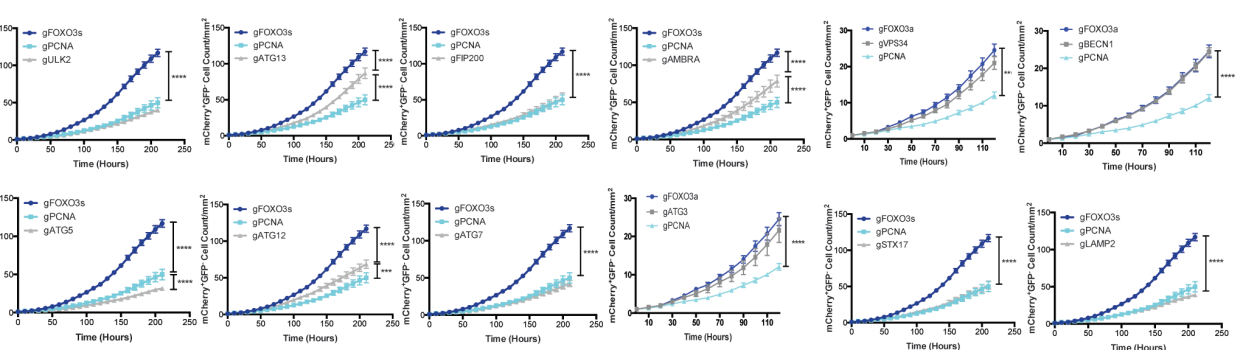
D

BT549



E

MAF-794



F

MDA-MB-468

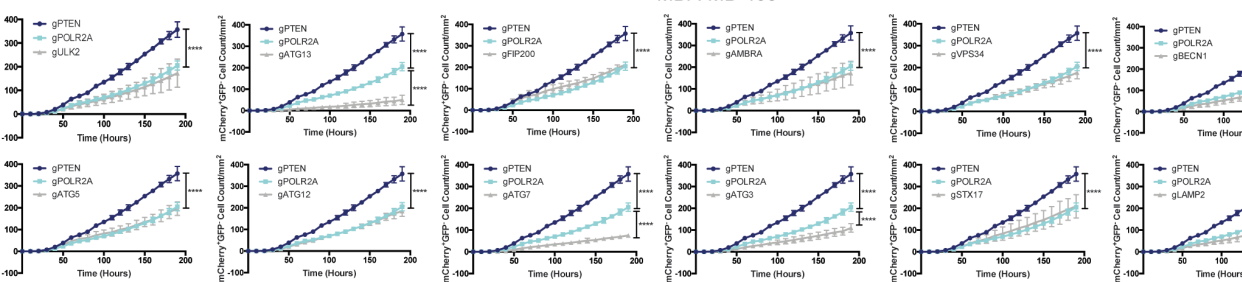


Figure S4, Related to Figure 2: Live-cell imaging CRISPR-RNP identifies autophagy dependent and independent cells. (A-F) Incucyte quantification of mCherry+/GFP- cell count after transfection with gRNAs targeting GFP and the indicated genes. The data are represented as mean \pm standard deviation (SD) for technical replicates (N of 2-3) and the graphs shown are representative of 2-3 individual experiments. Statistical analysis: 2-way ANOVA and the significance at the last time point is shown. * $p \leq 0.05$, ** $p \leq 0.01$, *** $p \leq 0.001$, **** $p \leq 0.0001$.

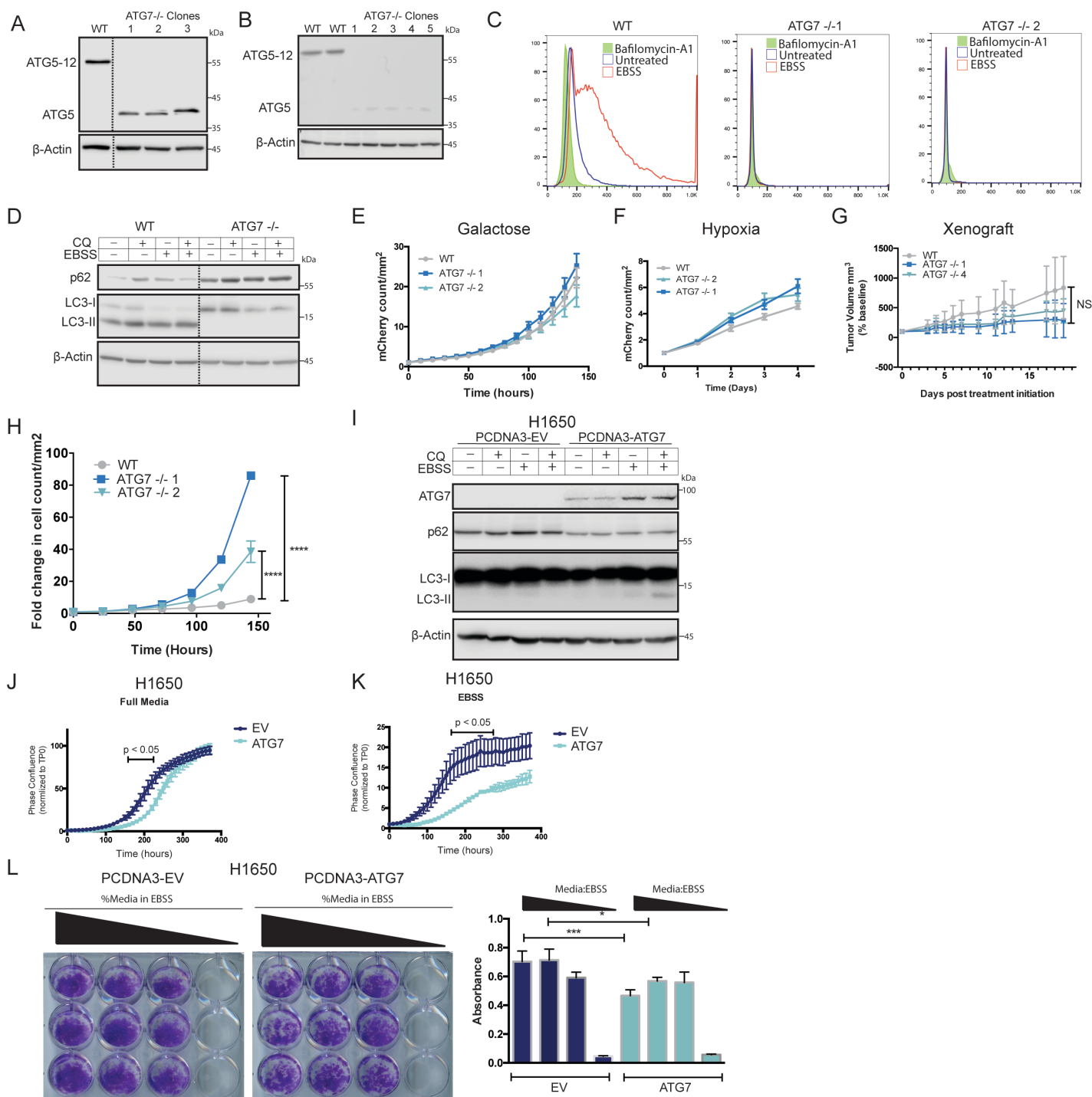


Figure S5, Related to Figures 3, 4: Autophagy-dependent cancer cells can undergo selection to circumvent complete inactivation of an autophagy regulator. (A-B) Western blot analysis in (A) BT549 and (B) H292 WT and ATG7^{-/-} clones. The blots shown are representative of 2 experiments. (C) The graphs show the shift in GFP expression in BT549 WT and ATG7^{-/-} clones with stable expression of LC3-mCherry-GFP 24hrs after starvation or Bafilomycin A1 (10nM). The data are representative of 2 experiments. (D) Western blot in BT549 WT and ATG7^{-/-} clones after 16hr starvation with or without Chloroquine (40 μ M) for 2hrs. The blots are representative of 3 experiments. (E-F) Incucyte live cell imaging mCherry+ cell counts over time in BT549 WT and ATG7^{-/-} clones grown in (E) media containing 10mM galactose or (F) hypoxia (1% O₂). The data are represented as mean \pm SEM for technical replicates (N of 3) and the graphs shown are representative of 3 individual experiments. (G) The mean tumor volume as a percentage of pre-treatment baseline, in vehicle treated mice bearing H292 WT or ATG7^{-/-} xenograft tumors. N= 5-7 mice/group. The data are represented as mean \pm SEM. Statistical analysis: 2-way ANOVA. (H) Incucyte quantification of mCherry+ cell count of H292 WT or ATG7^{-/-} clones after treatment with CQ (20 μ M). The data are represented as mean \pm SEM for technical replicates (N of 2-3) and the graphs shown are representative of 3 individual experiments. (I-L) NCIH1650 cells. (I) Western blot analysis in parental cells with stable expression of PCDNA3-EV or PCDNA3-ATG7. Representative of 3 experiments. (J-K) Incucyte growth assay monitoring phase confluency in (J) full growth media or (K) nutrient starvation with EBSS. The data are represented as mean \pm SEM for technical replicates (N of 3) and the graphs are representative of 2 experiments. Statistical analysis: 2-way ANOVA. (L) Left: Long term clonogenic growth assay in EV and ATG7 overexpressing cells treated with 100%, 75%, and 50% media in EBSS for 72 hours and then allowed to grow back in 100% media for 7 days. Right: Quantification of solubilized crystal violet from long term clonogenic growth assay. The data are represented as mean \pm SEM for technical replicates (N of 3). Statistical analysis: 2-way ANOVA. *p \leq 0.05, **p \leq 0.01, **** p \leq 0.0001

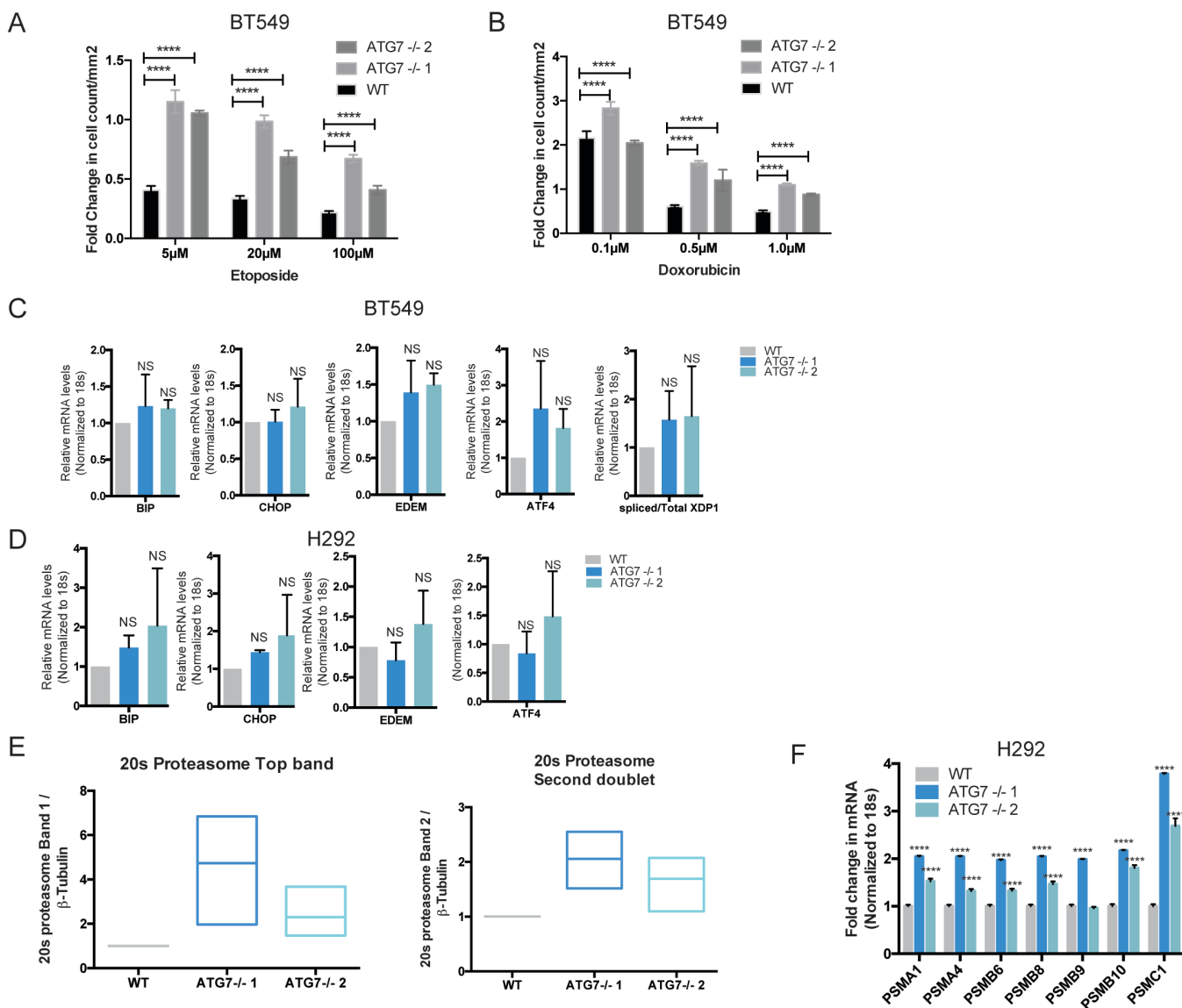


Figure S6, Related to Figure 5: Autophagy-dependent cancer cells are not more sensitive to DNA damaging agents and do not have increased ER stress but do have increased proteasomal subunits. (A-B) The Incucyte mCherry⁺ cell count/mm² calculated 4 days after treatment with (A) etoposide or (B) doxorubicin in BT549 WT or ATG7^{-/-} clones normalized to TP0. The data are represented as mean \pm SD for technical replicates (N of 3) and representative of 2 individual experiments. Statistical analysis: 2-way ANOVA. (C-D) qRT-PCR to measure mRNA levels of ER stress response genes relative to 18S mRNA levels in (C) BT549 and (D) H292 WT and ATG7^{-/-} clones. The data are represented as mean \pm SEM for experimental replicates (C: N of 3, D: N of 2). Statistical analysis: 1-way ANOVA. (E) Densitometric quantification of western blots probed for the 20S proteasome in Figure 5F for bands indicated with an asterisk. The data shown is from 3 independent experiments. (F) qRT-PCR to measure mRNA levels of 26S proteasomal subunits relative to 18S mRNA levels in H292 WT and ATG7^{-/-} clones. The data are represented as mean \pm SD for technical replicates (N of 2) and are representative of 2 individual experiments. Statistical analysis: 2-way ANOVA. **** p < 0.0001

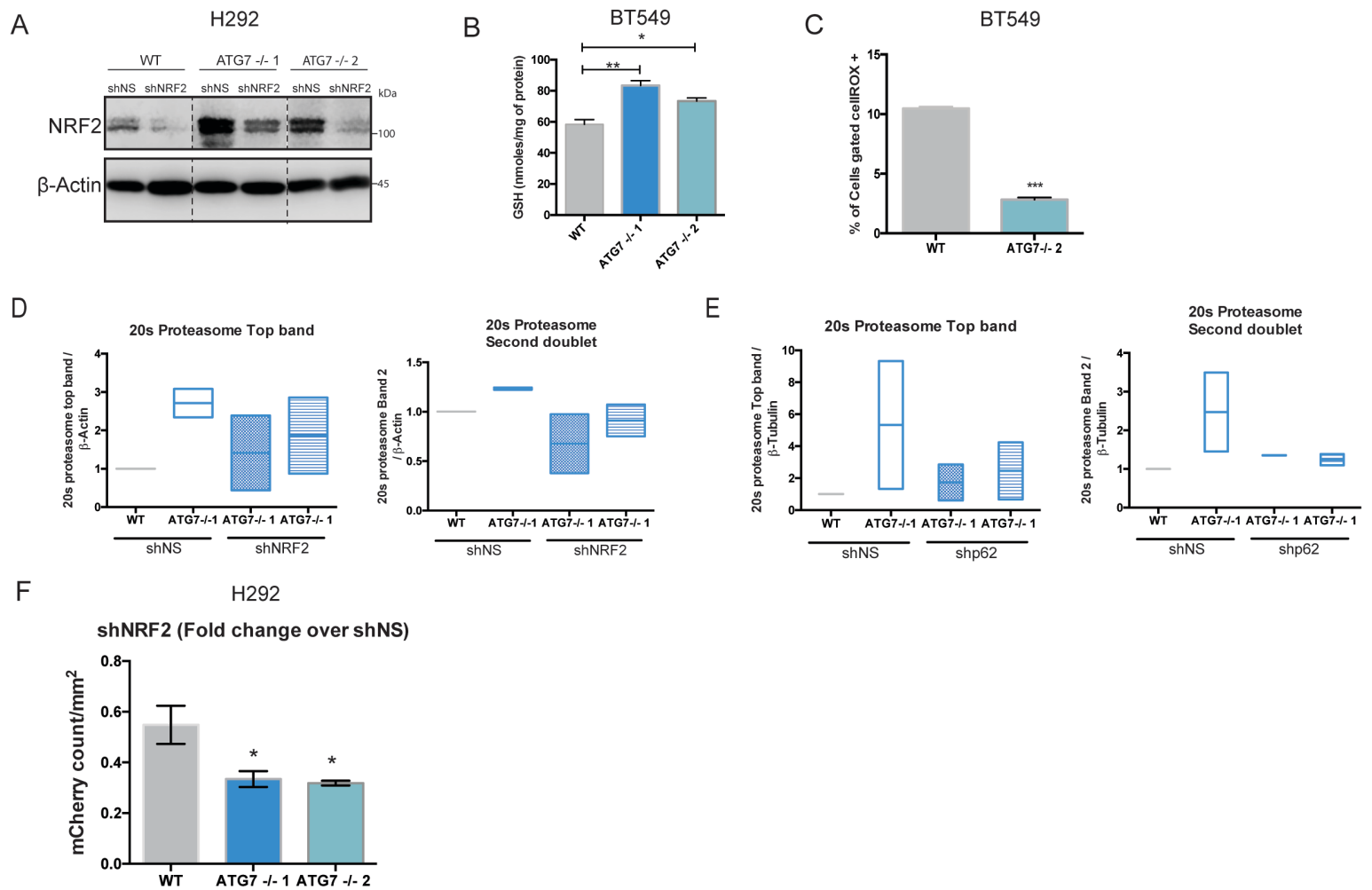


Figure S7, Related to Figures 6 and 7: ATG7^{-/-} clones have an increased dependency on NRF2. (A) Western blot analysis in H292 WT and ATG7^{-/-} clones with KD of NRF2. Dotted line indicates where unnecessary lanes were removed. The blots shown are representative of 2 experiments. (B-C) BT549 WT and ATG7^{-/-}. (B) Glutathione (GSH) levels were measured with a microtiter plate assay and normalized to total protein content. The data are represented as mean \pm SEM of three biological replicates. Statistical analysis: 1-way ANOVA. (C) Flow cytometry analysis of cellROX green was measured to identify levels of ROS. After gating live singlets based on forward and side scatter, a cellROX⁺ gate was drawn such that 10% of the untreated WT cells were counted as positive and used to quantify the percent of ATG7^{-/-} cells within that gate. The data is represented as the mean \pm SEM from two independent experiments. (D-E) Densitometric quantification of western blots probed for the 20s proteasome in (D) Figure 6B and (E) Figure 6D for bands indicated with an asterisk. The data shown is from 2 independent experiments. (F) KD of NRF2 induced effects on growth in H292 cells. The data are represented as mean \pm SEM of Incucyte mCherry⁺ cell count/mm² 9 days after cells were transduced with shNRF2 and normalized to shNS for technical replicates (N of 3). The data are representative of 2 individual experiments. Statistical analysis: 1-way ANOVA. *p \leq 0.05, *** p \leq 0.001